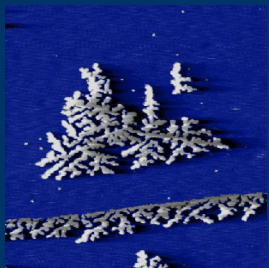


Biomolecular Tubes and Fibers

Alexander Bittner

"Chemical Nanostructuring and Self-Assembly"



Abt. Kern - Nanoscale Science

Max Planck Institute for Solid State Research, Stuttgart, DE



Group "Self-Assembly"

CIC Nanogune Consolider, Donostia-San Sebastian, ES

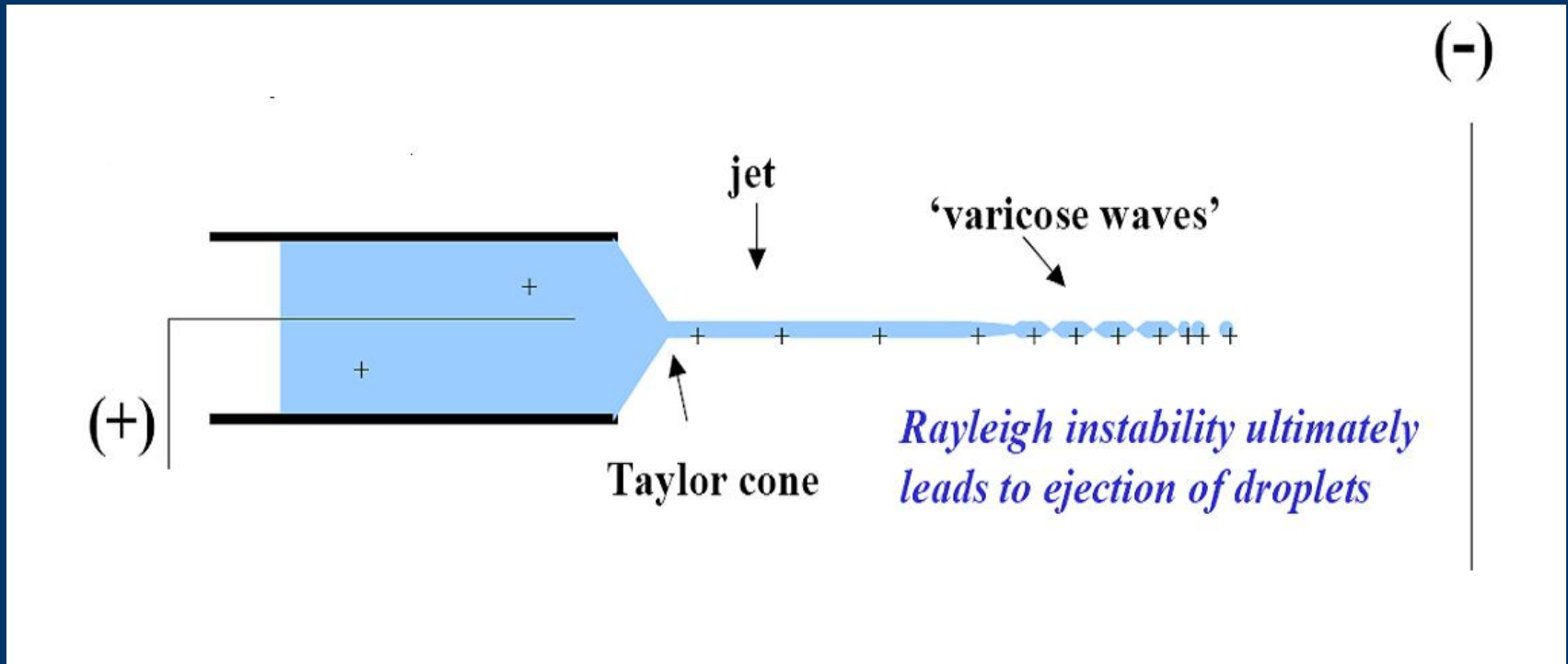


Electrospun self-assembling peptides

Nikola Malinowski
Sebastian Loscher
Gurvinder Singh
Darya Amoli
Marten Tolk
Guillaume Hupin

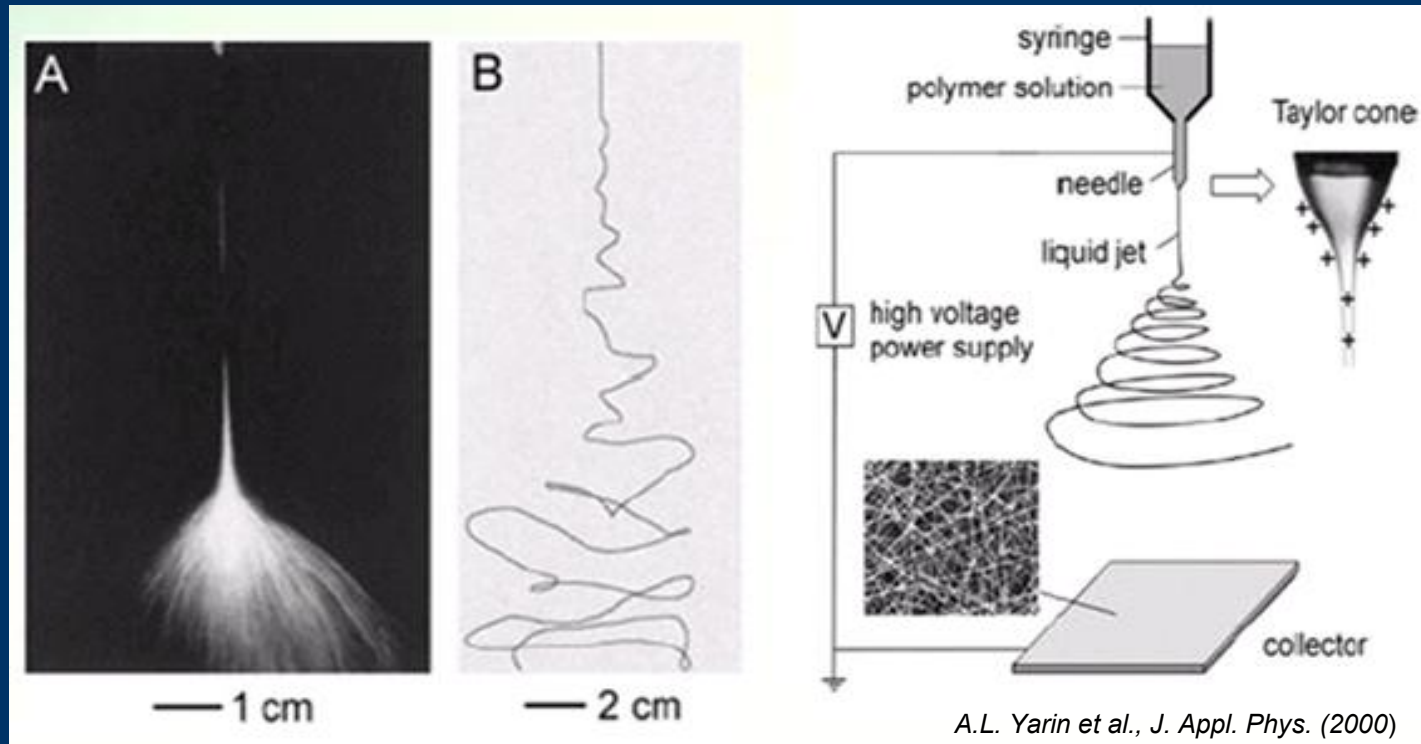
(MPI-FKF Stuttgart etc.)

Electrospraying



Surface tension $\sigma/r \leftrightarrow E^2$ field (charges on the jet)

Electrospinning (of polymers)

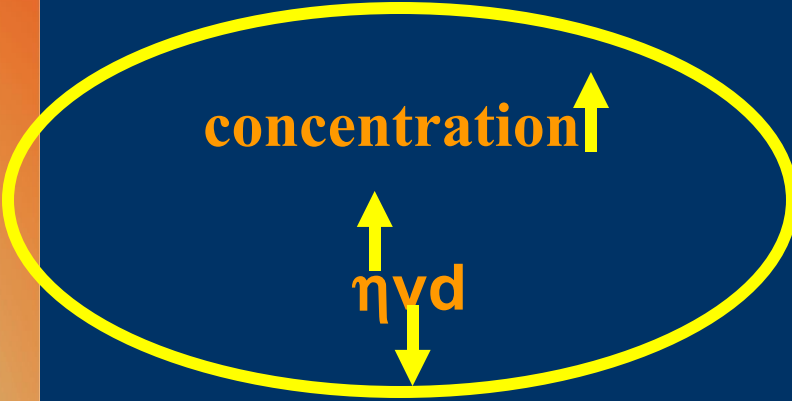
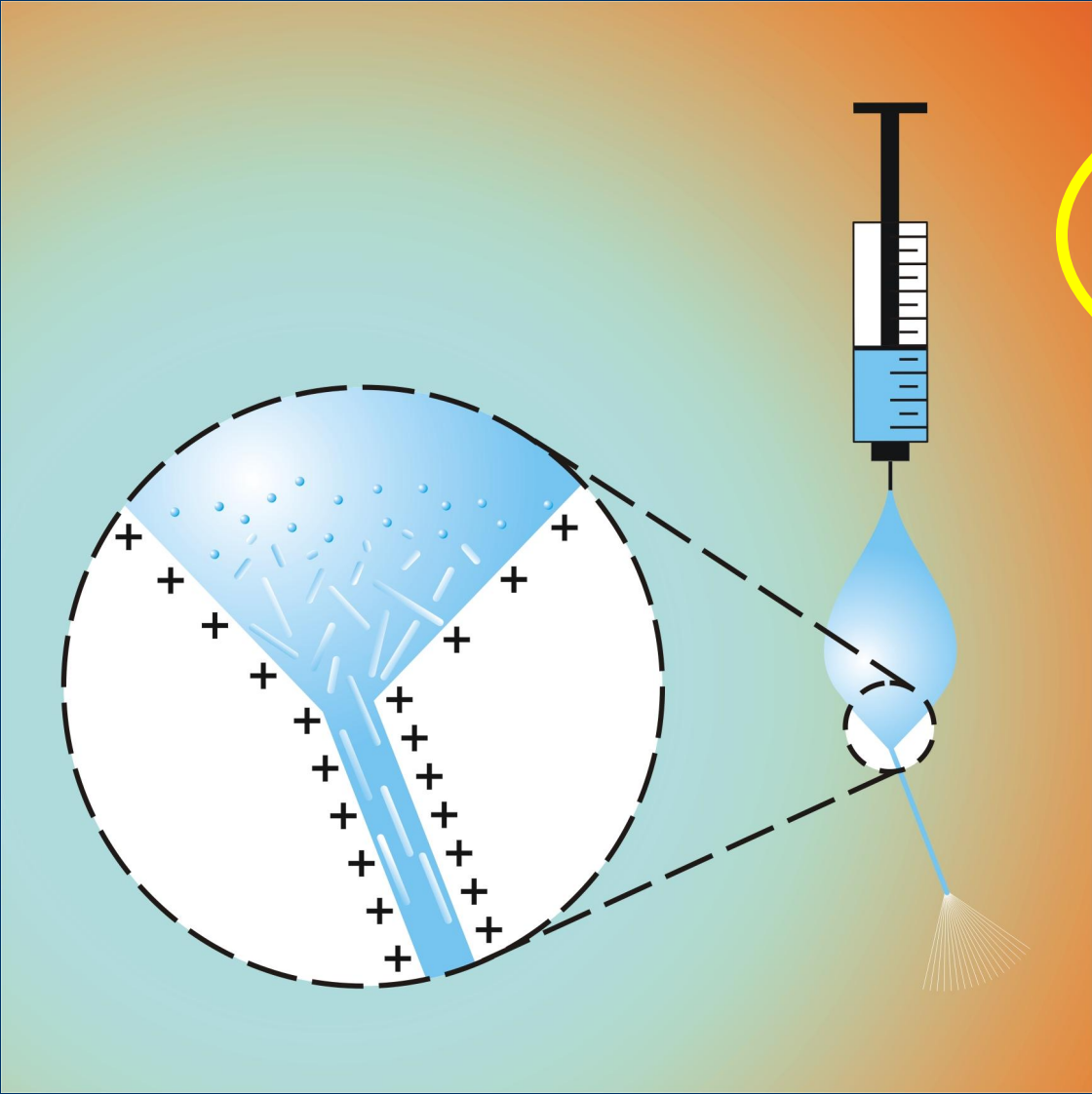


Highly viscous solution of polymers; constant feed; kV potential

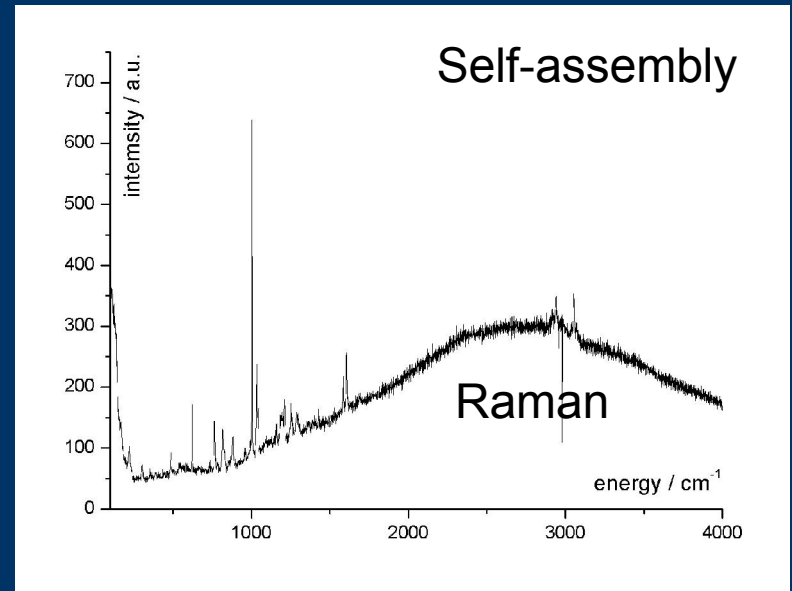
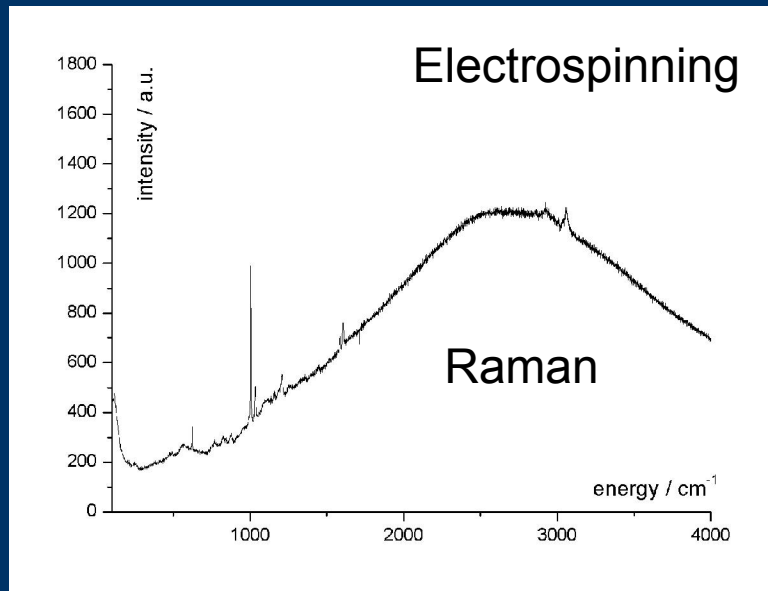
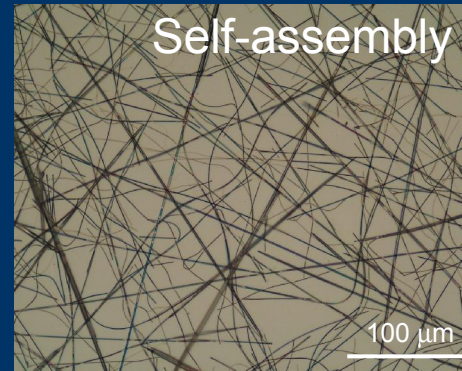
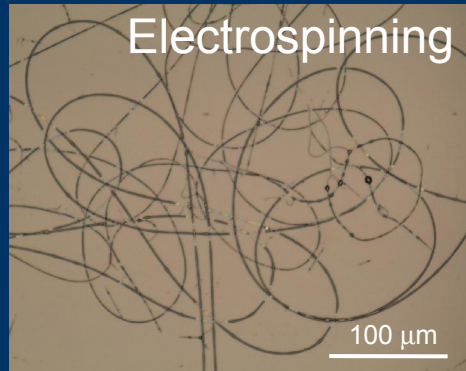
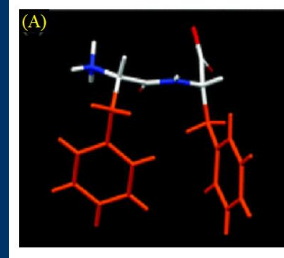
; Viscosity η changes with radius r ! But: high η and v for cone.

Evaporation of solvent: Concentration \uparrow Vapour pressure \uparrow

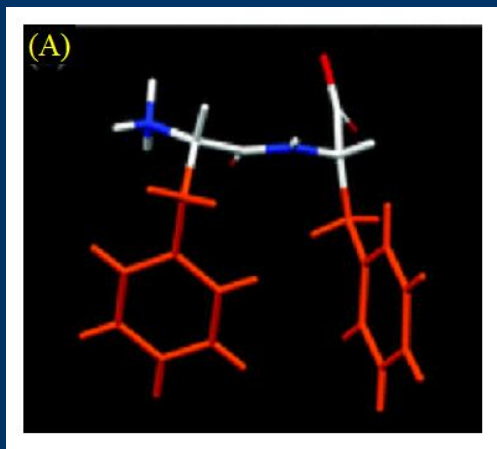
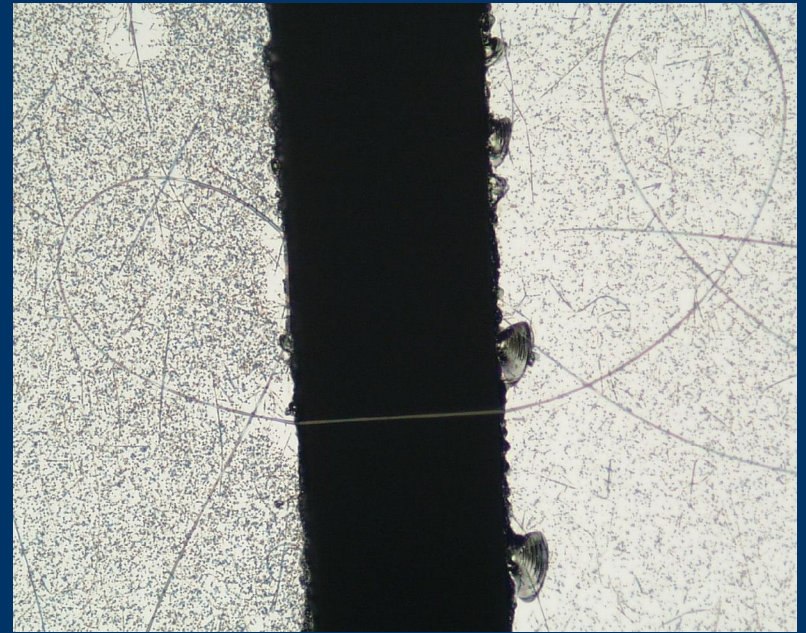
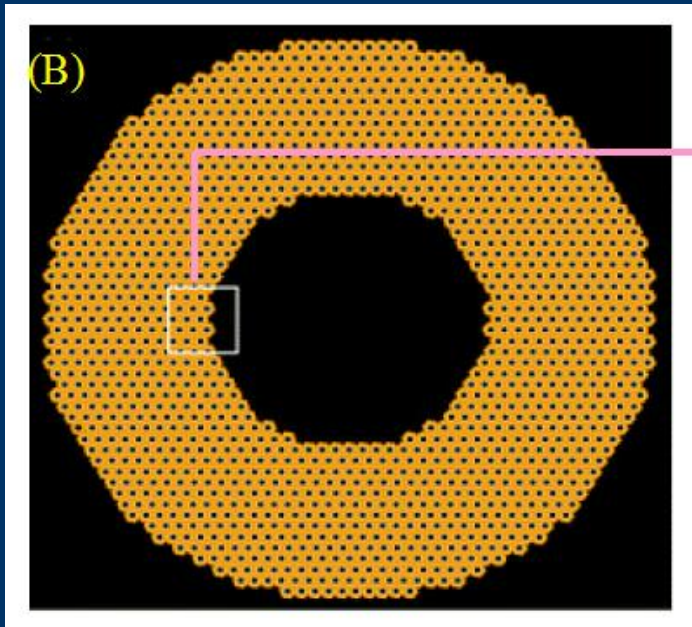
Electrospinning of *monomers*



Electrospinning of self-assembling di-phenylalanine (Phe-Phe, FF)



Diphenylalanine tubes



Stretching and bridging over 0.1 mm gaps

The Tobacco mosaic virus in nanoscale science

Former Bittner group (Dept. Klaus Kern)

Gabriel Baralia, Sinan Balci, Kei Noda, Mato Knez

Stuttgart University, Dept. of Mol. Biology & Plant Virology (Holger Jeske)

Anan Kadri, Anna Müller, Christina Wege, Emil Ruff

Max Planck Institutes Stuttgart

MPI-MF: StEM (electron microscopy group)

MPI-FKF: Von Klitzing group

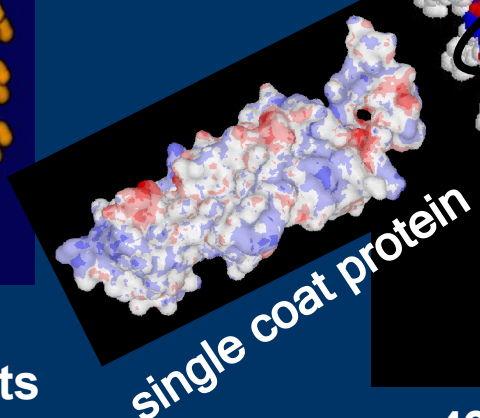
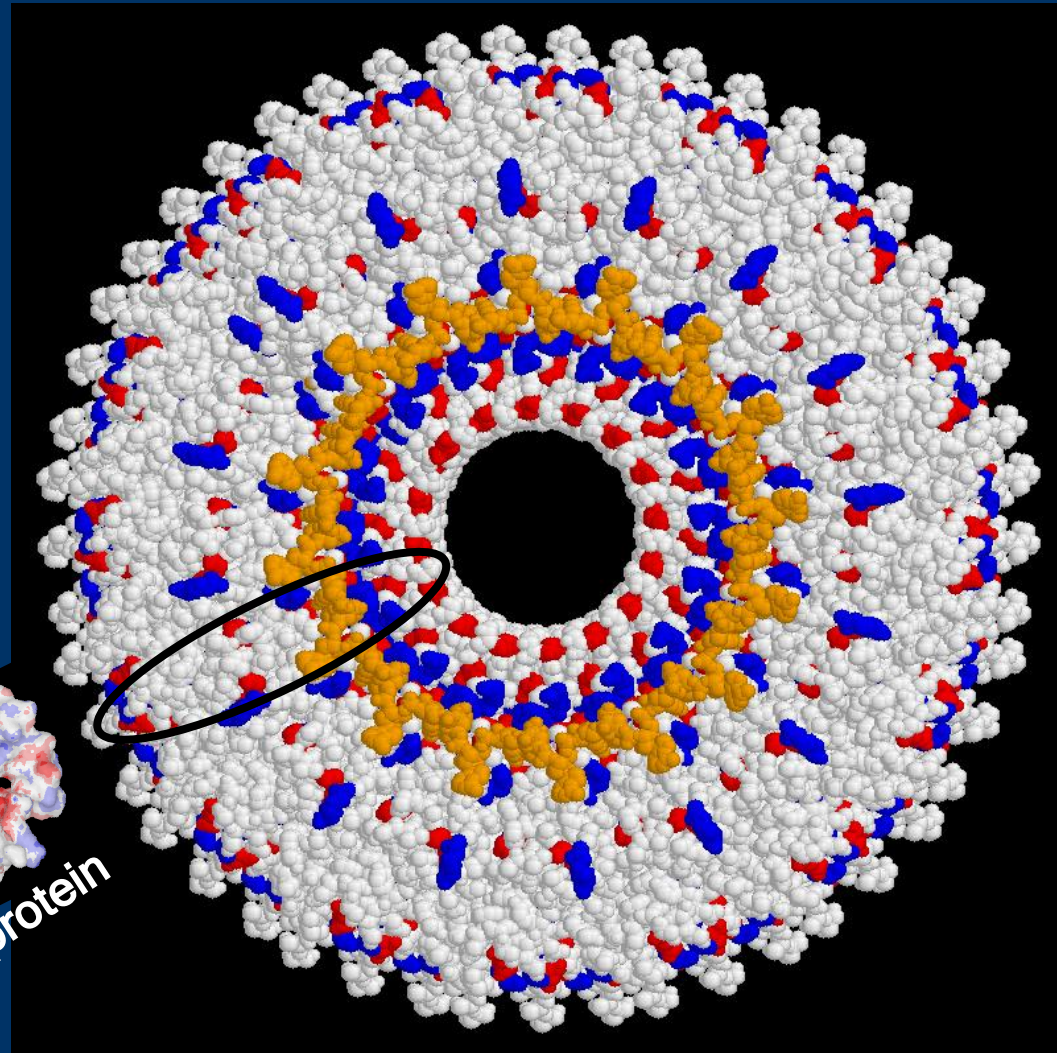
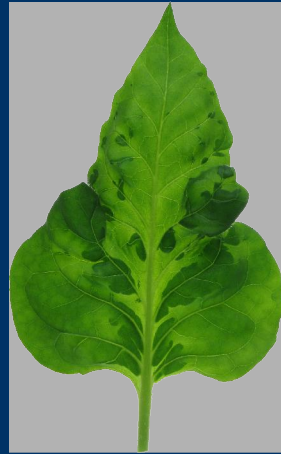
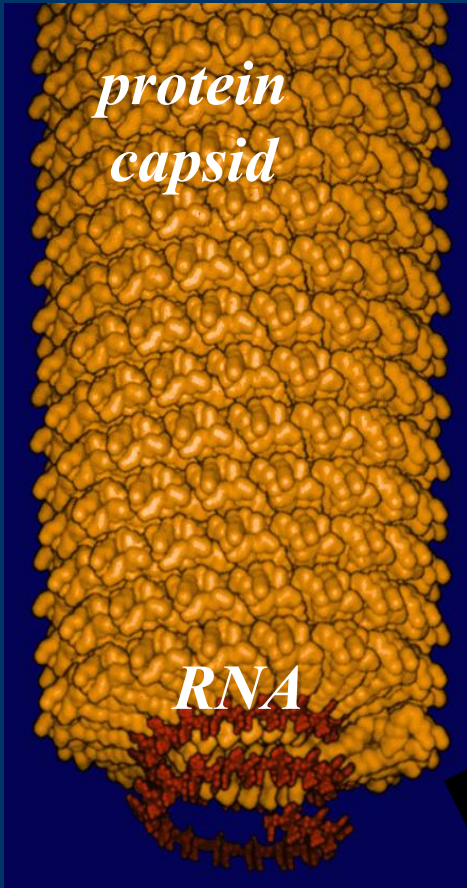
Ulm University (Carl Krill III)

Zhenyu Wu, Chenchen Ma

Financing:

Max-Planck-Gesellschaft
Deutsche Forschungsgemeinschaft
Alexander von Humboldt-Gesellschaft
Kompetenznetz "Funktionelle Nanostrukturen" Baden-Württemberg

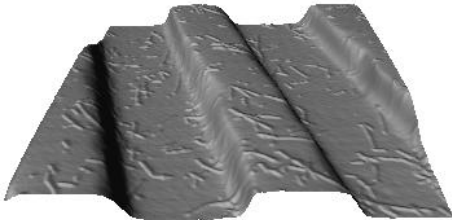
Tobacco mosaic virus (TMV)



viral rod (partial)
2100 protein subunits
300 nm length

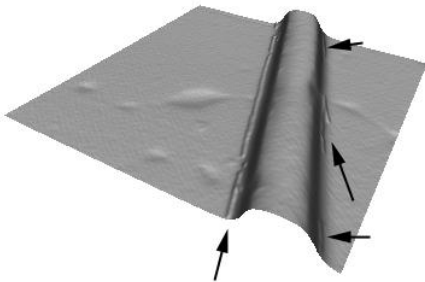
"Edge printing" of virus lines

Poly(dimethylsiloxane) stamp,
O₂ plasma-treated

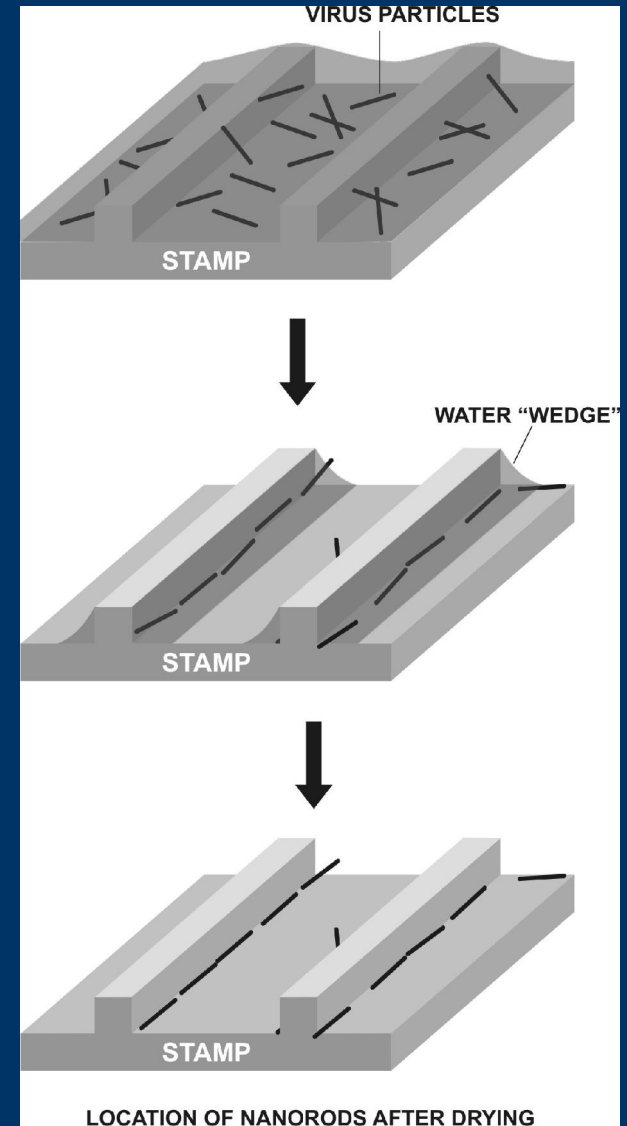


**TMV adsorbed on
stamp**

Poly(dimethylsiloxane) stamp,
O₂ plasma-treated

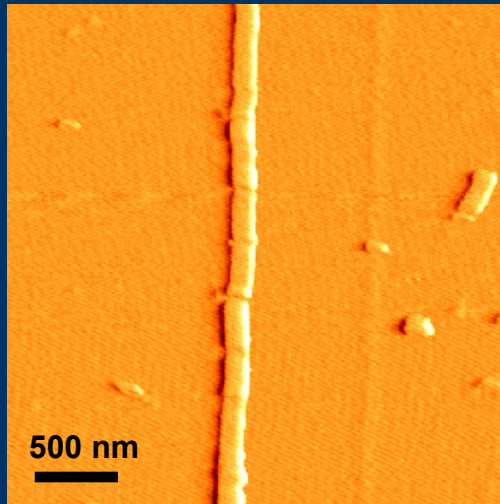


**TMV adsorbed on
stamp, low conc.,
blow dried →
discontinuous
dewetting**



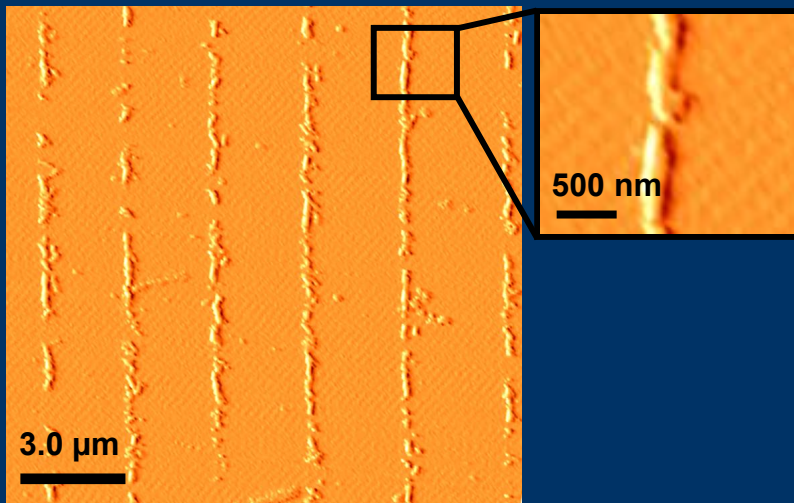
"Edge printing" of virus lines

Oxidized Si

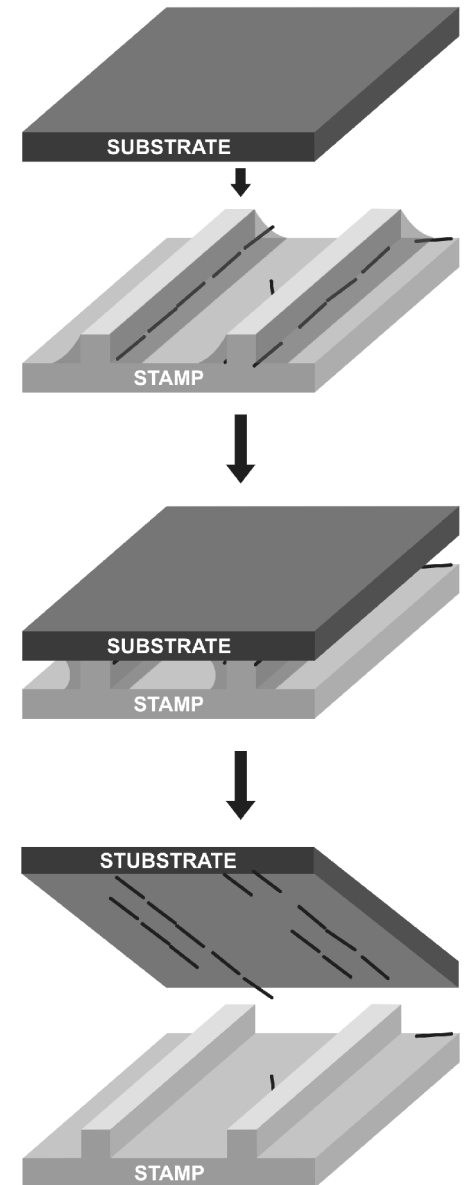


TMV lines printed on oxidized silicon wafer; width < 30 nm

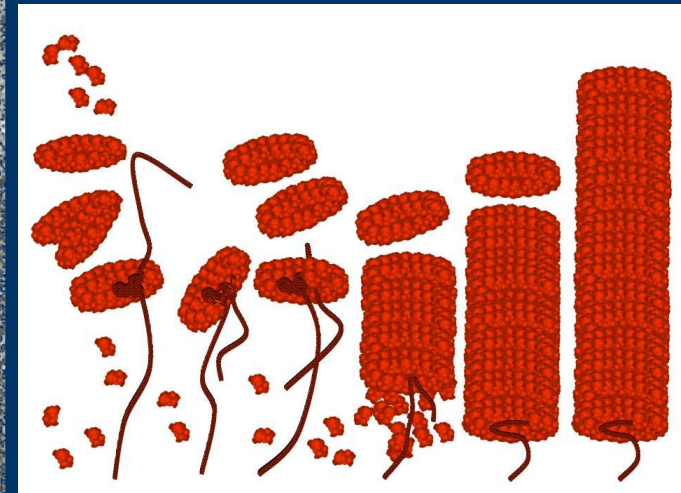
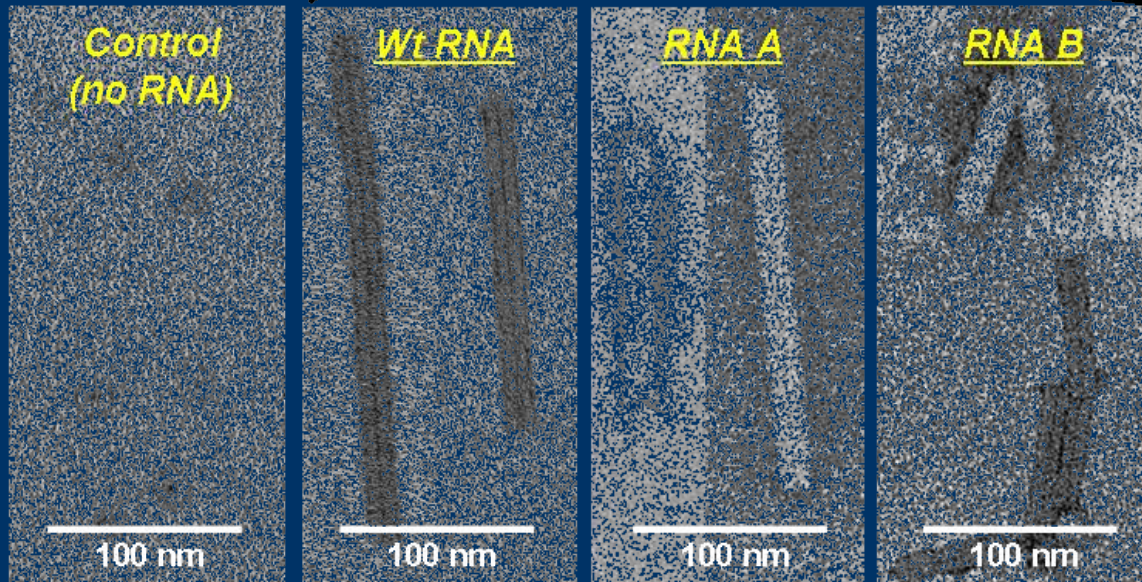
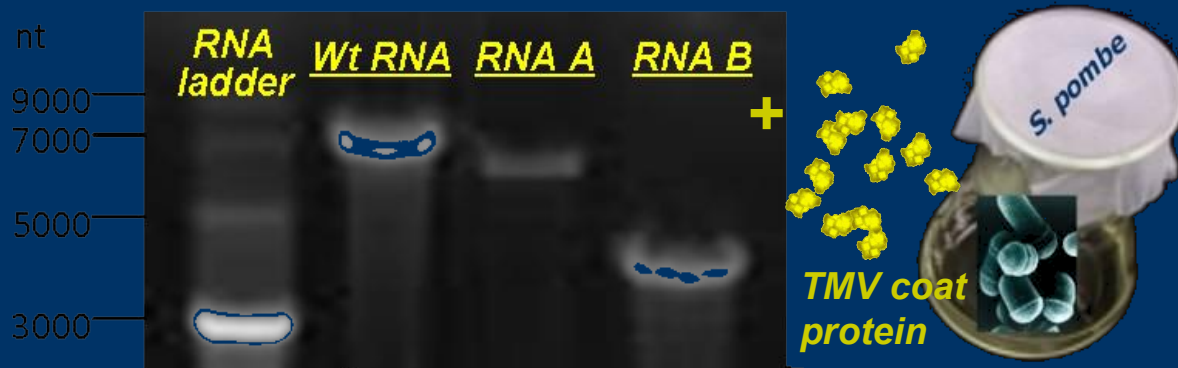
Oxidized Si



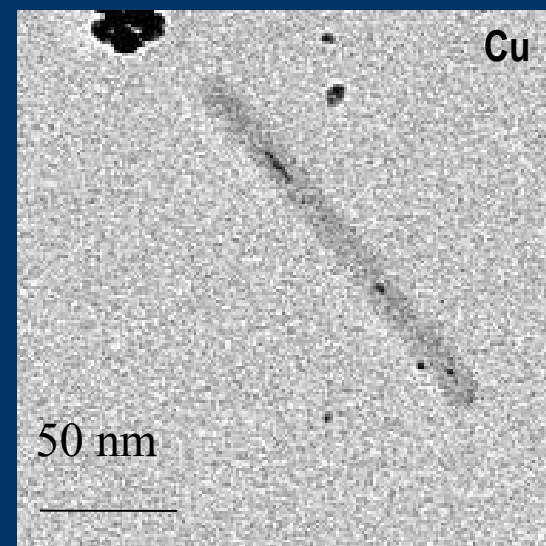
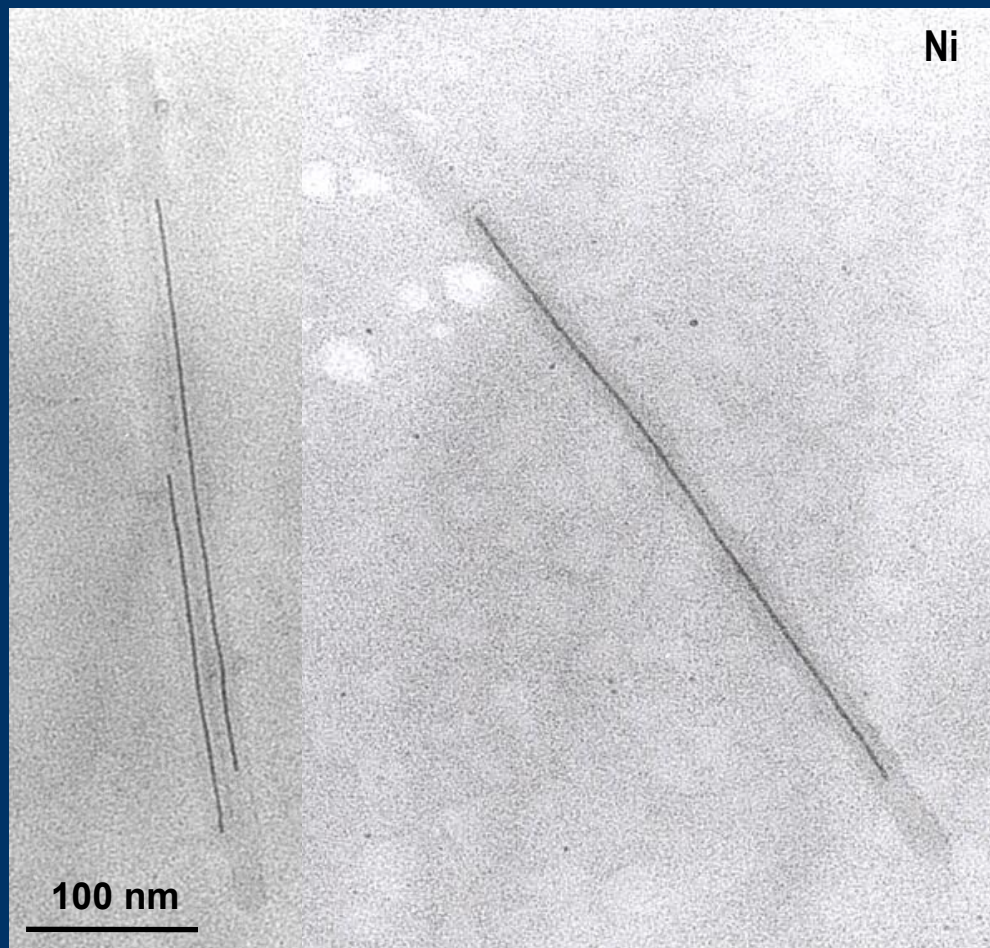
MODEL OF THE PRINTING PROCESS



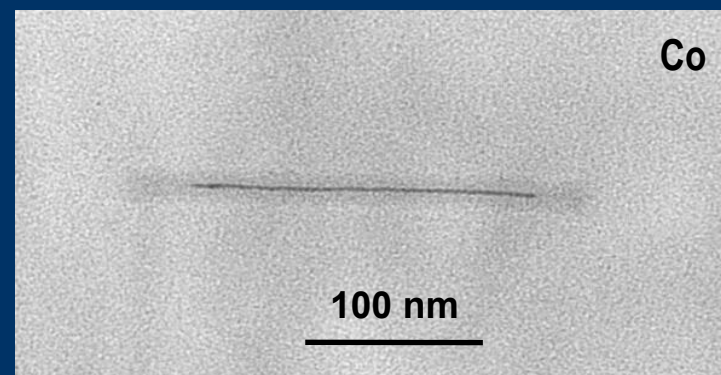
Assembly of "artificial TMV" = coat protein + RNA



The “true” nanoscale: 3 nm wires in TMV

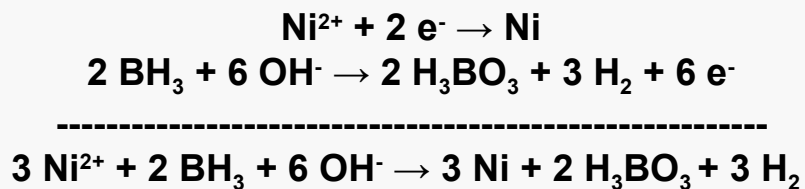


Pd/Cu
deposition;
wires and
clusters



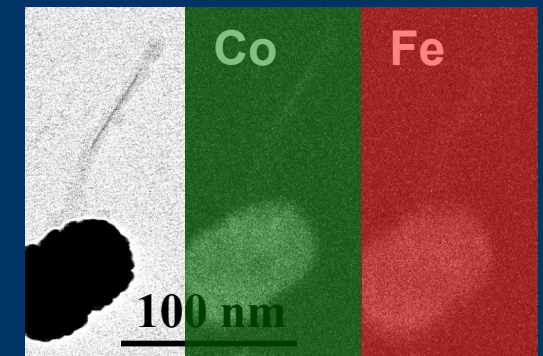
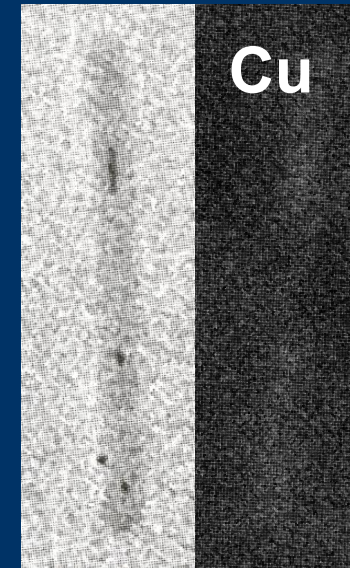
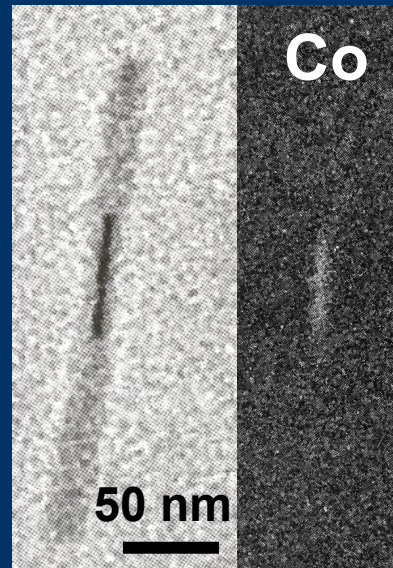
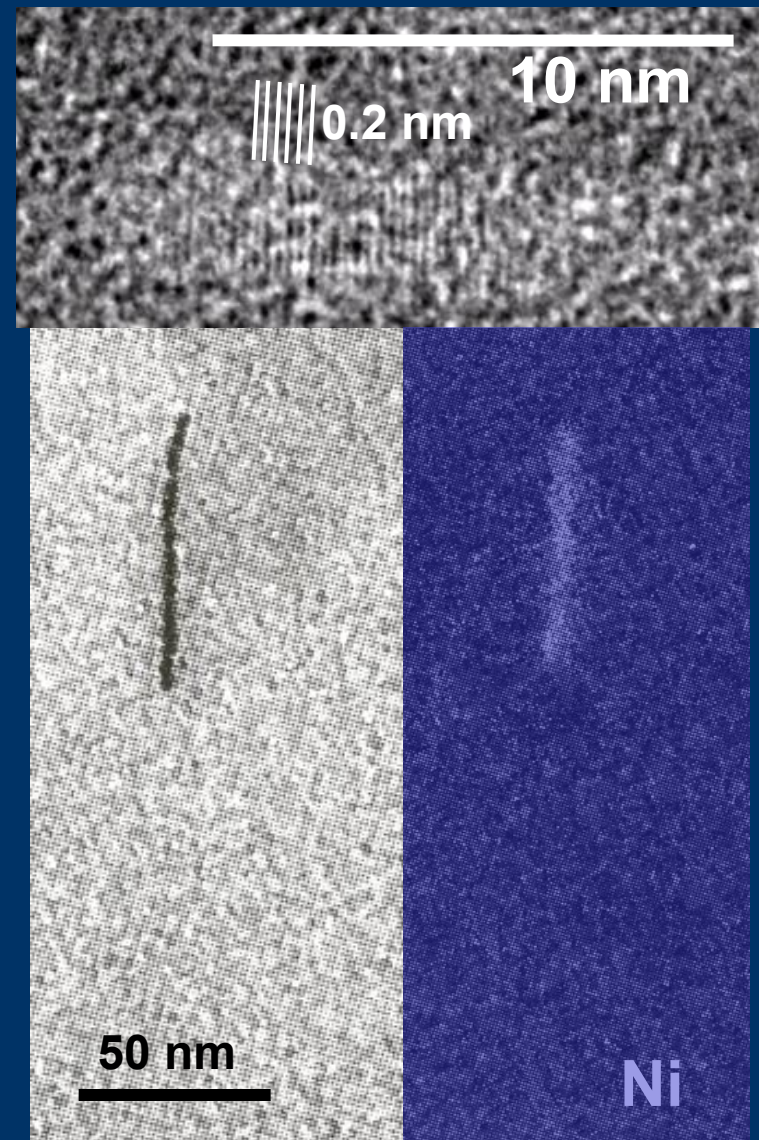
TEM after Pd(II) activation and Ni deposition

Pd/Co deposition

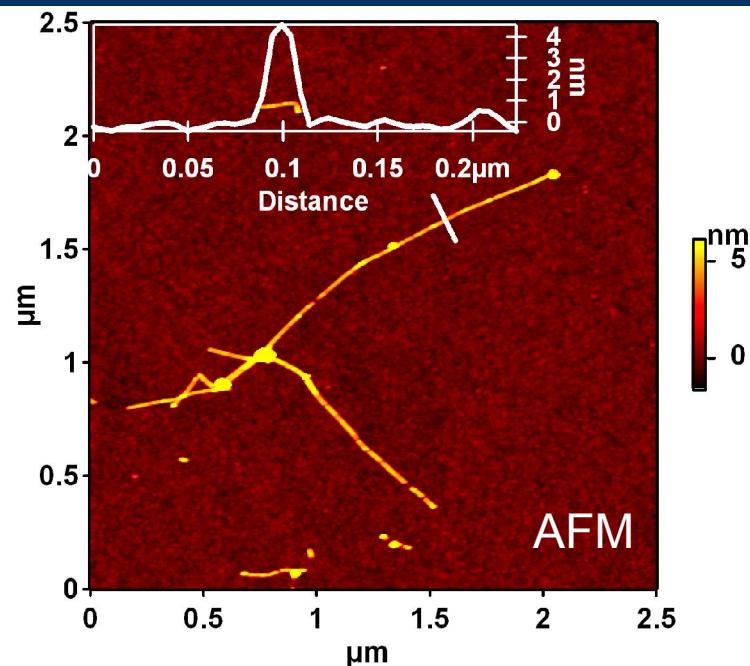
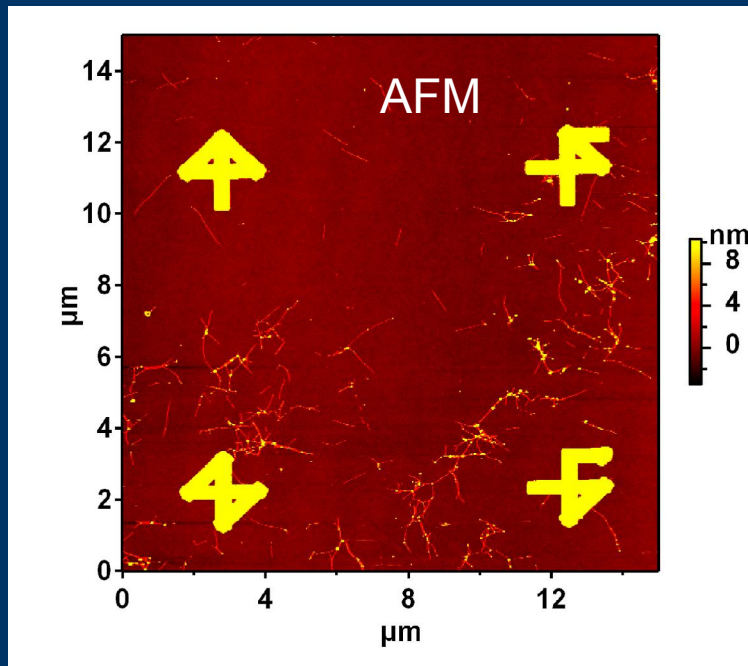
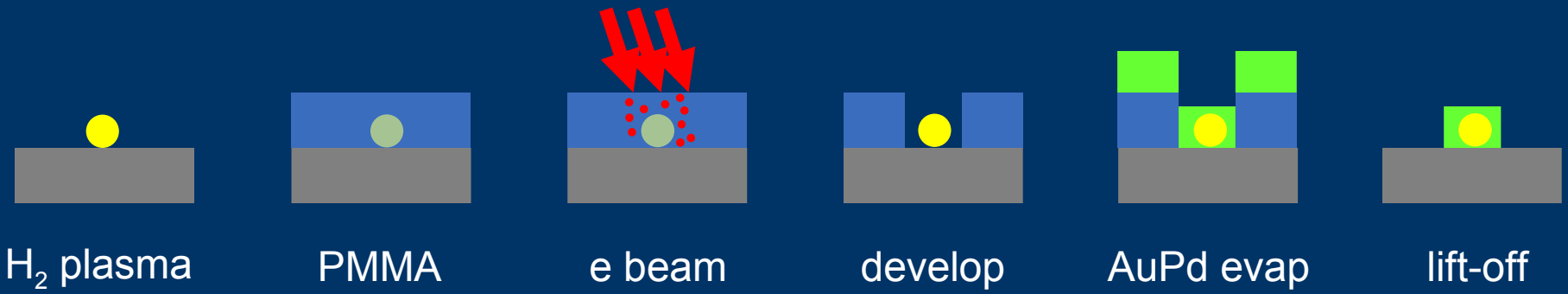


Energy filtering TEM of 3nm wires in virions

Chemical and structural analysis on the sub-5nm scale: Pure metal (little O); for Ni oriented crystallites, [111] in wire axis

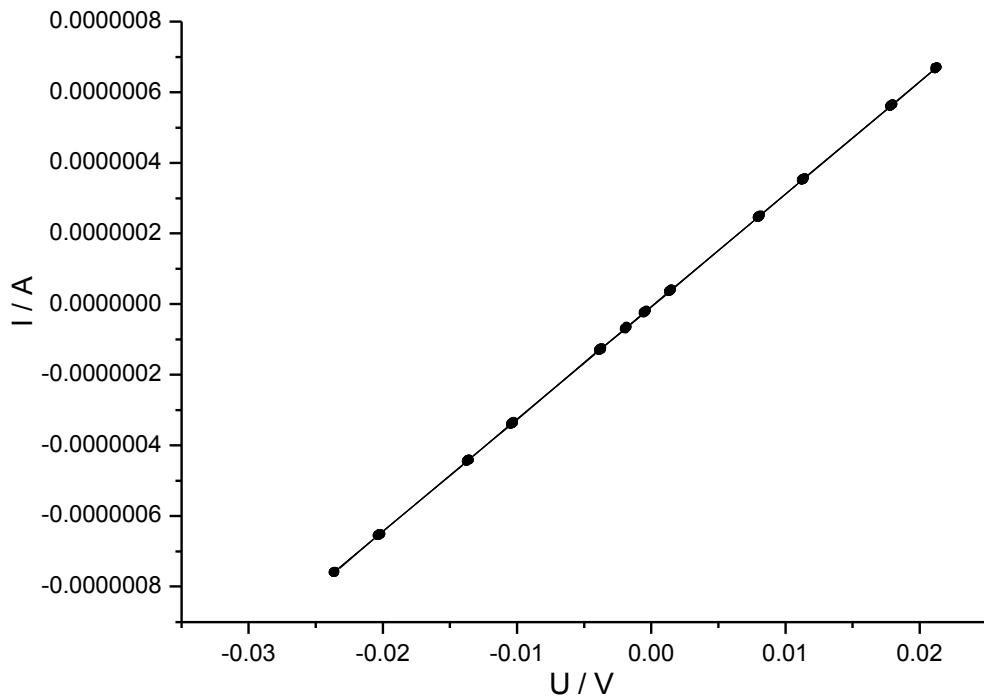


Lithography for contacting

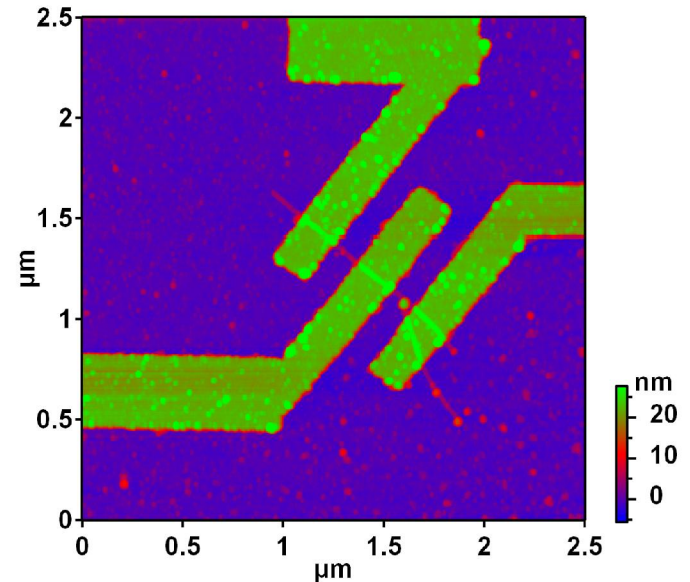


Extremely long Ni wires in E50Q-TMV on silicon / silicon oxide wafers with markers

The first contacted 3 nm nickel wires



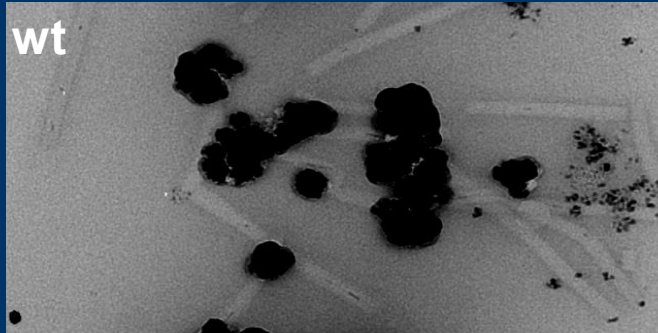
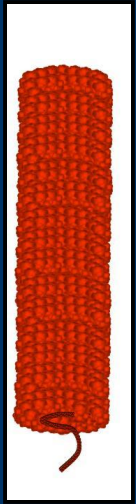
Current-voltage curve



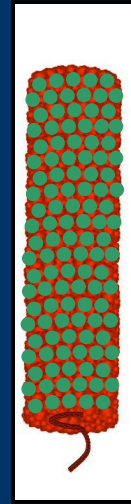
AFM topography

Extremely long Ni wires in virus-like TMV coat proteins
Removal of all organic material by oxygen/hydrogen plasma
Electron beam lithography, AuPd contacts
Ca. 20 k Ω

Mutations and phosphate control the deposition

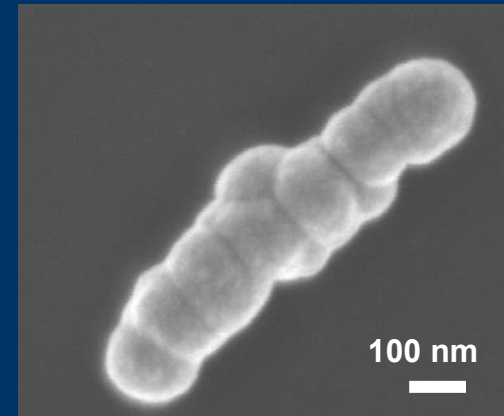


Pd(II) sensitization and Ni(II) electroless deposition



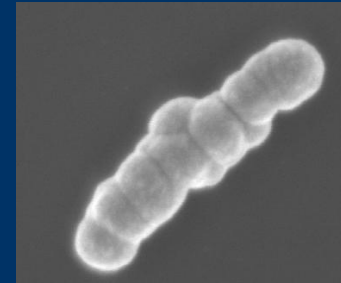
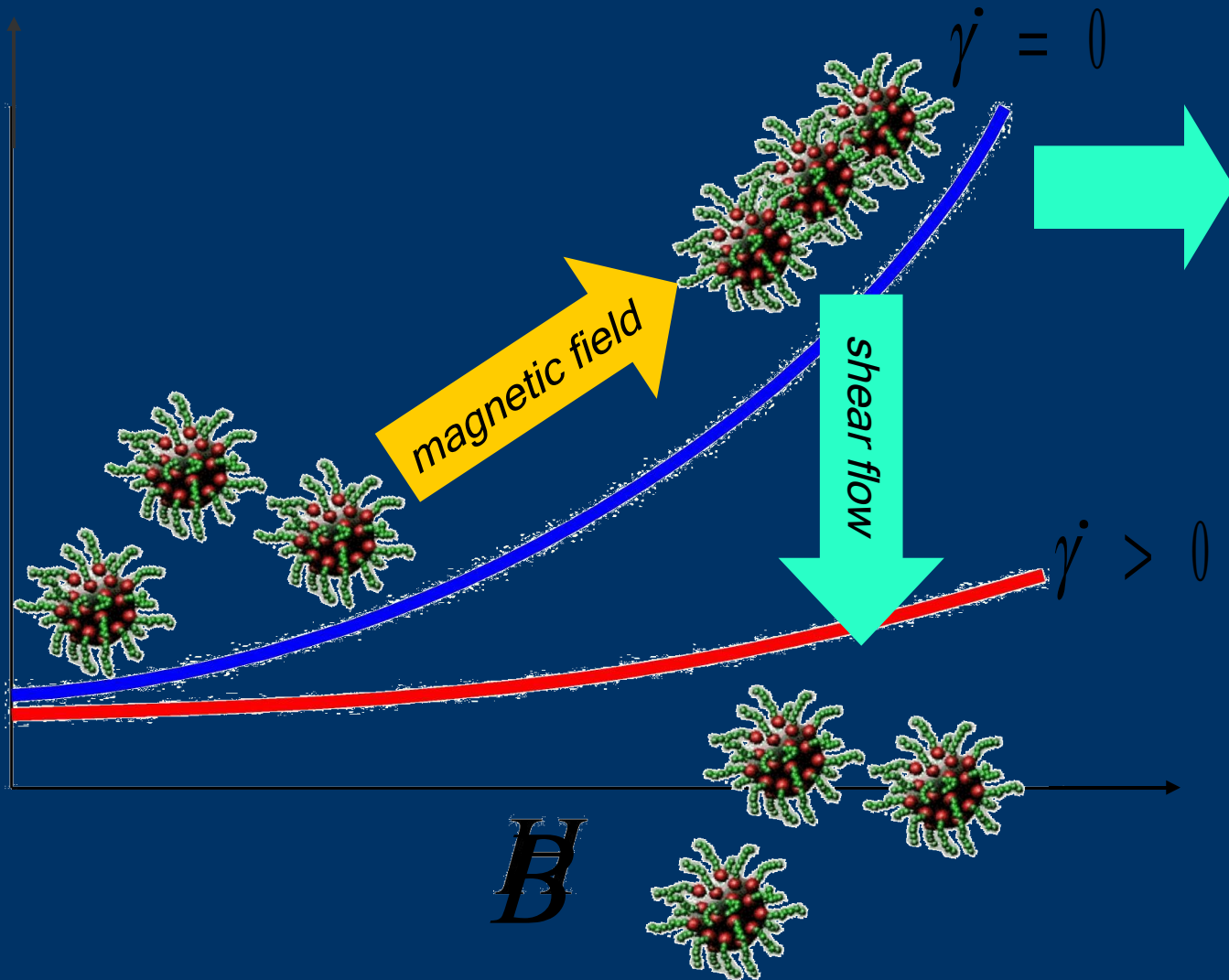
6·2100 His residues, Pd(II) and Ni(II) coordination: deposition on the coat

Pd(II) sensitization with phosphate traces:
Ni(II) electroless deposition on coat

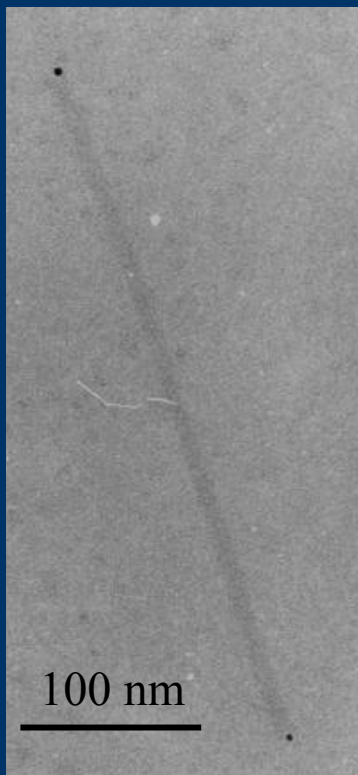


Ferrofluids: Shear thinning

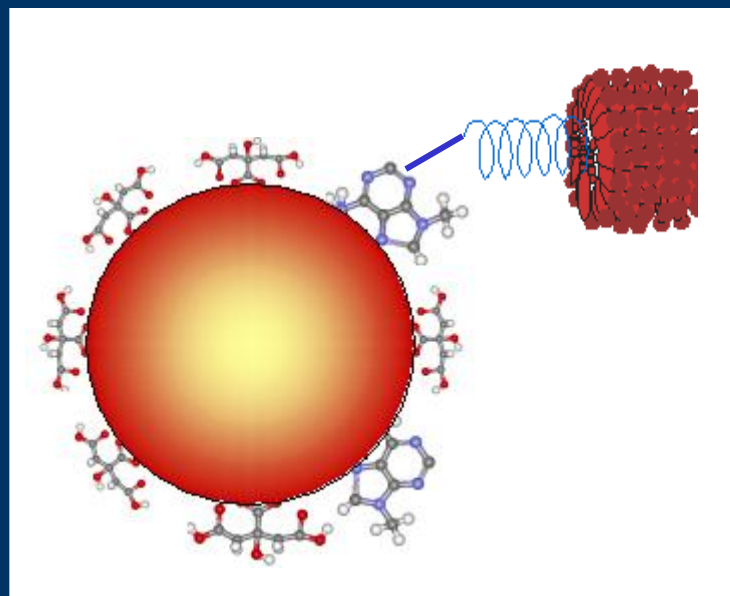
Viscosity



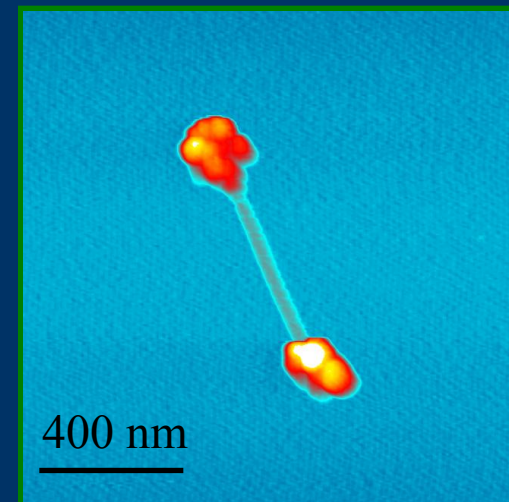
Deposition at the ends: Metal dumbbells



6 nm gold clusters



Citrate-covered gold binds to RNA (freed by the gold particle)



AFM: 50 nm gold nodules by “enhancement”, electroless deposition of gold on gold

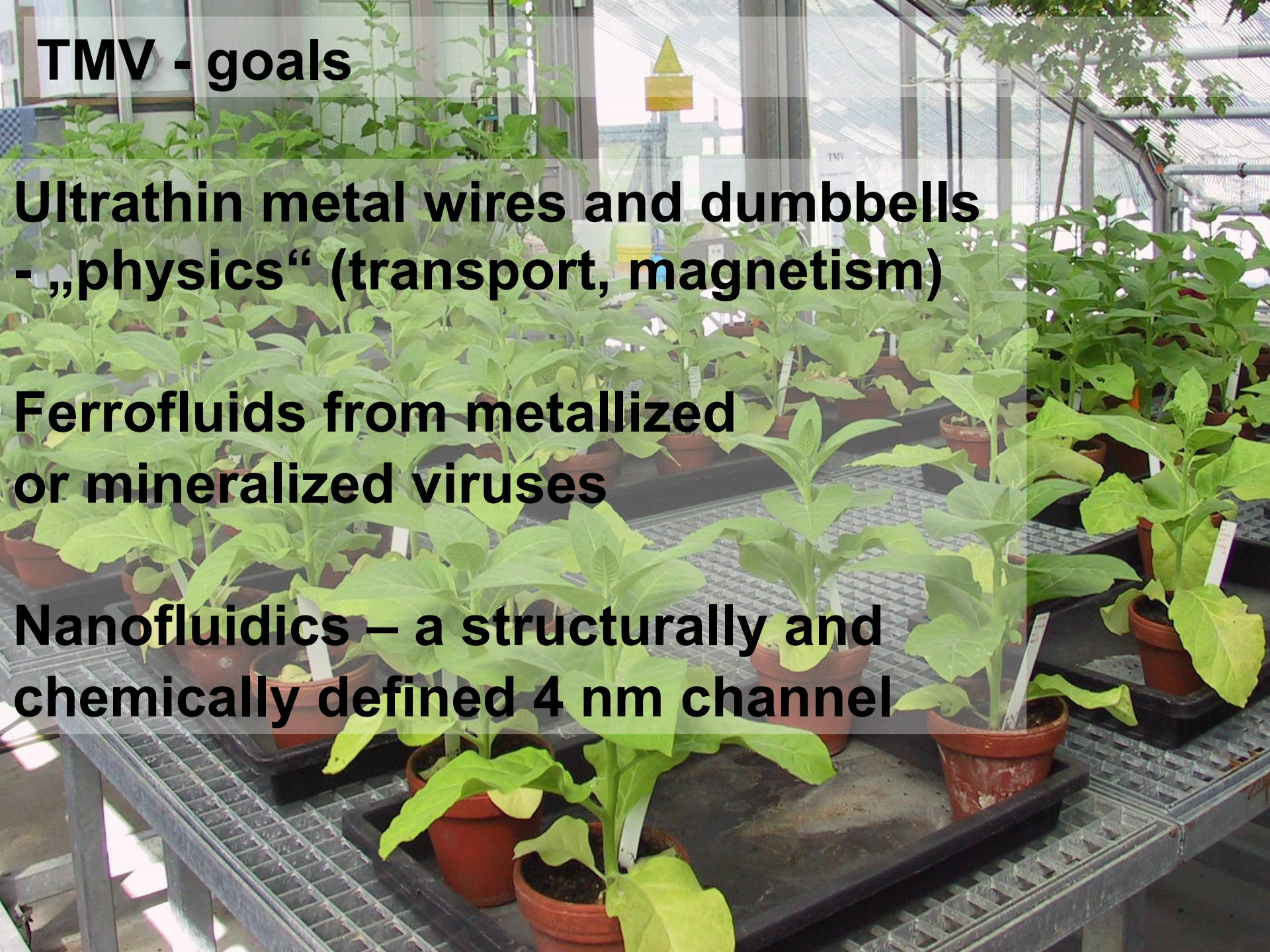
**Complete self-assembly of complex structures;
switchable containers?**

TMV - goals

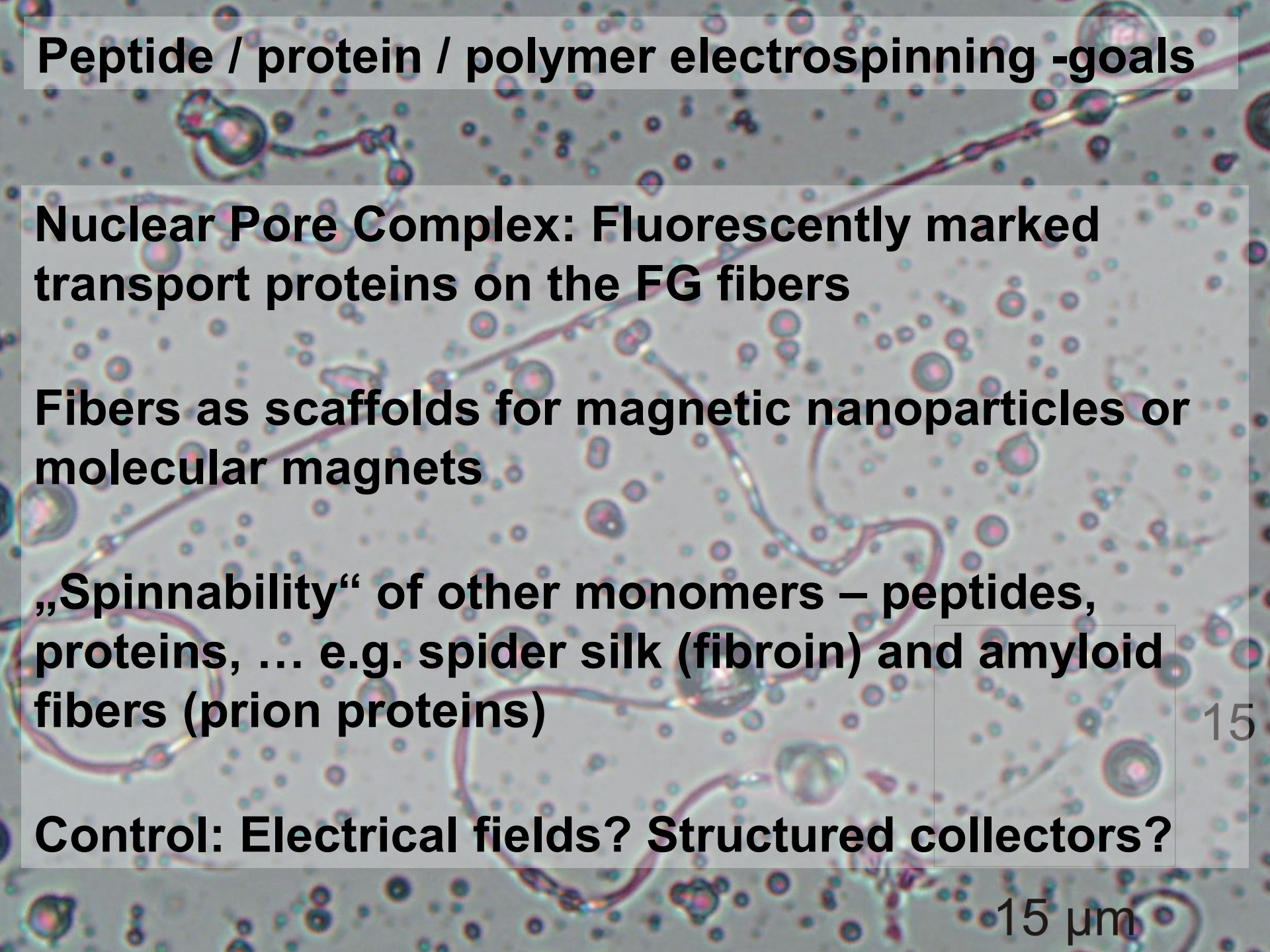
**Ultrathin metal wires and dumbbells
- „physics“ (transport, magnetism)**

**Ferrofluids from metallized
or mineralized viruses**

**Nanofluidics – a structurally and
chemically defined 4 nm channel**



Peptide / protein / polymer electrospinning -goals

A micrograph showing a network of thin, interconnected fibers. The fibers are primarily purple and blue, with some segments appearing yellow or green. Numerous small, bright, circular spots are scattered throughout the fibers and the background, representing fluorescently marked transport proteins. The overall structure is complex and porous.

Nuclear Pore Complex: Fluorescently marked transport proteins on the FG fibers

Fibers as scaffolds for magnetic nanoparticles or molecular magnets

„Spinnability“ of other monomers – peptides, proteins, ... e.g. spider silk (fibroin) and amyloid fibers (prion proteins)

Control: Electrical fields? Structured collectors?

15 μm

15

nanoGUNE

Director:
J.-M. Pitarke

Nanomagnetism



Berger
Vavassori

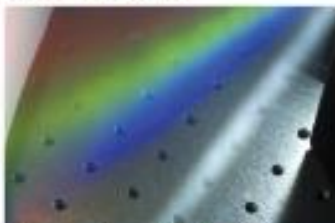
Design, fabrication and characterization of nanomagnetic and spintronic structures and devices.

Nanodevices



Nanofabrication of devices and its impact on nanobiotechnology, nanomagnetism and nanomechanics.

Nanooptics



Hillen-
brand

Advanced near-field optical microscopy. Nanophotonic structures and devices.

Nanoscale Imaging



Scanning probe microscopy. Electron microscopy.

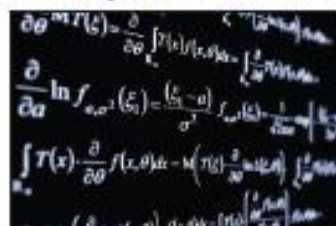
Self-assembly



Bittner

Synthesis, functionalization and processing of nanomaterials. Self-assembly of complex structures.

Theory & Simulation



Theoretical methods and computational tools for the study of the nanoscale.

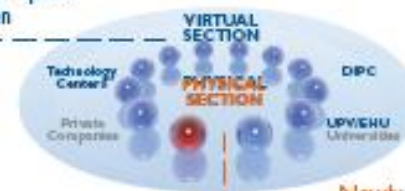
Nanobiotechnology



Nabiev

Biofunctional nanoparticles, non-bio/bio interfaces, and nanobioassemblies.

Open collaborative research space between



Newly created with its own personnel, building and facilities