## **Impact of agglomeration on the relaxometric properties of gadolinium oxide nanoparticles as a contrast agent for MRI**

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**Context:** Magnetic resonance imaging (MRI) is a non invasive biomedical imaging modality that allows high resolution diagnostics. The signal in MRI is provided by relaxing  ${}^{1}H$  protons. In order to increase the efficiency of tissue differentiation, it is often necessary to increase the signal in specific organs or tissues. To date, contrast agents are used in 30% of all clinical scans and the most used are gadolinium chelates [1]. These chelates are referred to as "positive- $T_i$ " contrast agents since they enhance the signal from relaxing  ${}^{1}H$  protons. In the context of cellular imaging however, those chelates do not allow the study of cell migration in vivo since they are not efficiently retained within the cells. This is a niche application for which ultrasmall gadolinium oxide nanoparticles (US-Gd<sub>2</sub>O<sub>3</sub>,  $\varnothing$  core = 3 nm) have been considered [2, 3]. Nanoparticles can be efficiently ingested and retained by cells, leading to improved contrast with  $T_1$ -weighted MRI sequences [4, 5]. However, once internalised by the cells, the nanoparticles tend to agglomerate in endosomes [4]. The present study aimed at evaluating the impact of agglomeration on the relaxometric properties of  $Gd<sub>2</sub>O<sub>3</sub>$  nanoparticles. In order to avoid interference with organic materials, here only aqueous suspensions of nanoparticles were characterized (without cells).

**Materials and Methods:** US-Gd<sub>2</sub>O<sub>3</sub> were synthesized by hydrolysis in a polyol solvent [6, 7]. As-synthesized nanoparticles are covered with diethylene glycol (DEG-Gd<sub>2</sub>O<sub>3</sub>). Then, they were dialyzed against water. Due to the presence of contaminating DEG, the resulting nanoparticle suspensions tend to form nanoagglomerates of hydrodynamic size ranging from 3 nm (individual nanoparticles) to about 105 nm. The hydrodynamic radius of agglomerates was studied by dynamic light scattering (DLS), while longitudinal relaxivities  $(r_1)$  were measured on a Stelar field cycling relaxometer (NMRD) from 0.01 to 10 MHz. The relaxometric study was completed by using dedicated relaxometers (Bruker Minispec, 10, 20, 60 MHz) to measure <sup>1</sup>H longitudinal and transversal relaxation times  $(T_1$  and  $T_2)$  at clinical fields. High resolution NMR spectrometers were used to characterize the suspensions at 300 and 500 MHz (high-field MRI). Gd concentration was measured by ICP-MS.

**Results and conclusions:** Agglomeration of DEG-Gd<sub>2</sub>O<sub>3</sub> results in a slight decrease of both  $r_1$ and r2. However, even 105 nm agglomerates still perform well as "positive-*T1*" contrast agents, as suggested by  $r_2/r_1$  ratios close to 1.5 at 60 MHz, compared to 1.3 for individual nanoparticles. The simulated signal intensity is 10.5% higher for individual nanoparticles. At clinical fields (∼1.5 T, 60 MHz), NMRD curves indicate a promising maximum in  $r_1$  relaxivity. This maximum occurs at magnetic fields six times higher than for individual ultra-small iron oxide nanoparticles (USPIOs). This result suggests that  $Gd_2O_3$  nanoparticles are more suitable than USPIOs to provide positive contrast in clinical 1.5 to 3 T MRI [8, 9]. DEG-Gd<sub>2</sub>O<sub>3</sub> could also be used in high-field pre-clinical MRI (at 4-7 T), a range of magnetic fields for which USPIOs cannot be used to provide positive contrast because the  $T_2^*$  effects become too important [8].

## **References:**

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## **Figures:**



**Figure 1:** Longitudinal  $(R_1)$  and transversal  $(R_2)$  relaxation rates of DEG-Gd<sub>2</sub>O<sub>3</sub> agglomerates







