

## Comparison of the results of the microbiologic quality of an untreated water sample using conventional culture media and a DNA chip for simultaneous detection of microorganisms

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# INTRODUCTION

Water quality assessment  
(drinking and recreational)



Public Health

Legislation / EU Directives  
98/83/CE and 2006/7/EC

Presence of pathogenic  
organisms

Conventional  
microbiological methods

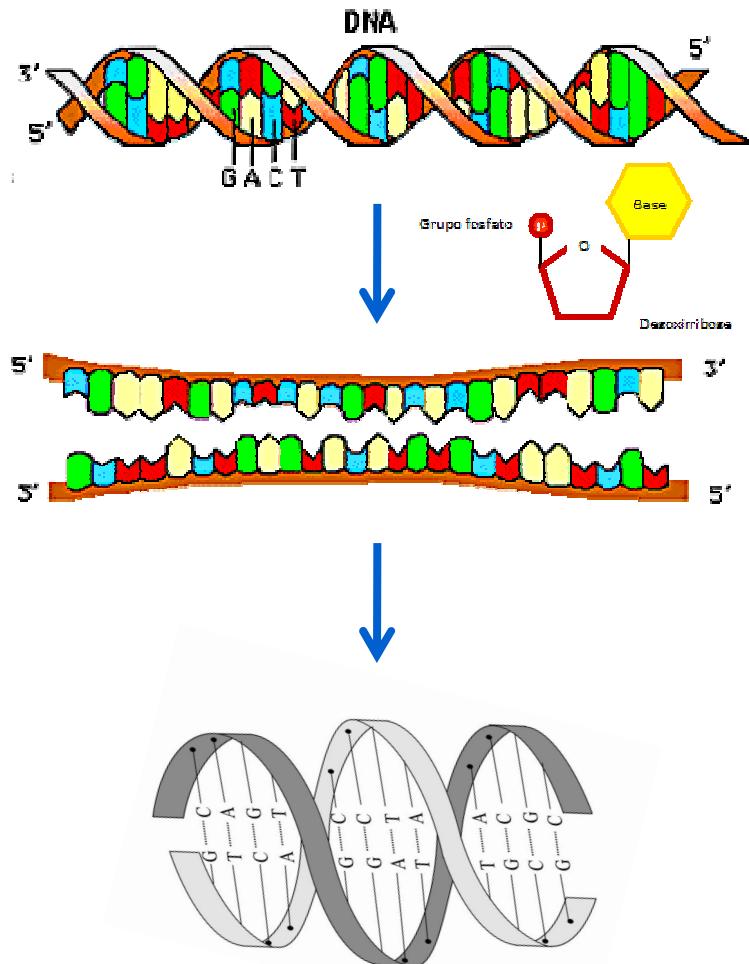
Time consuming, indicator bacteria  
limitations...

## Goal of AQUACHIP project

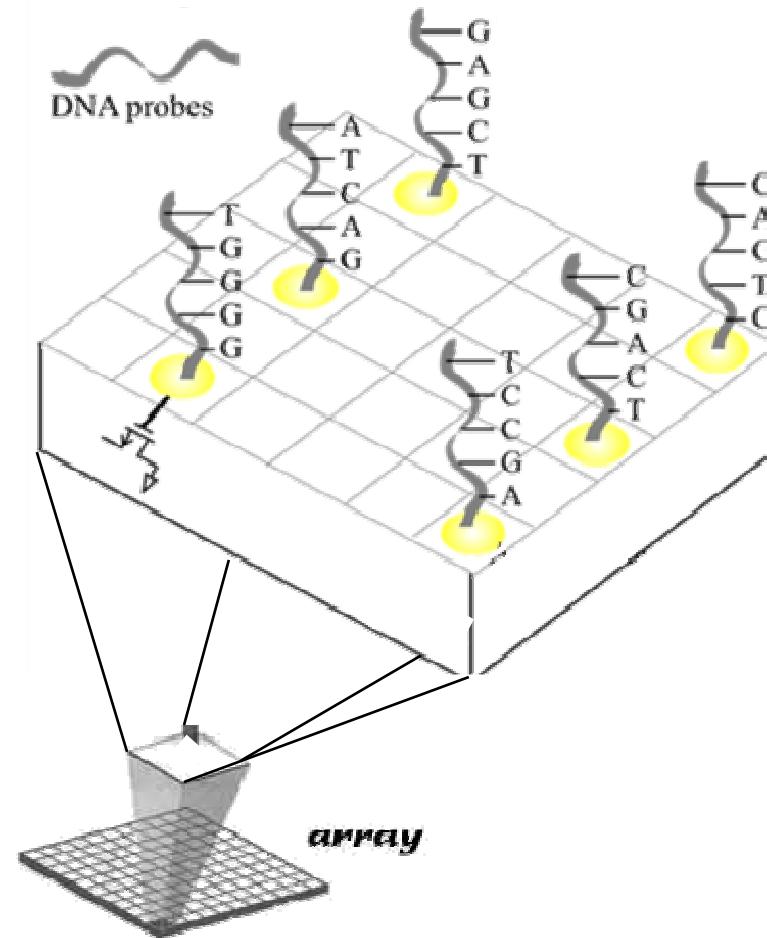
Development of a rapid method for simultaneous detection of several microorganisms in water samples by the use of a DNA chip

2

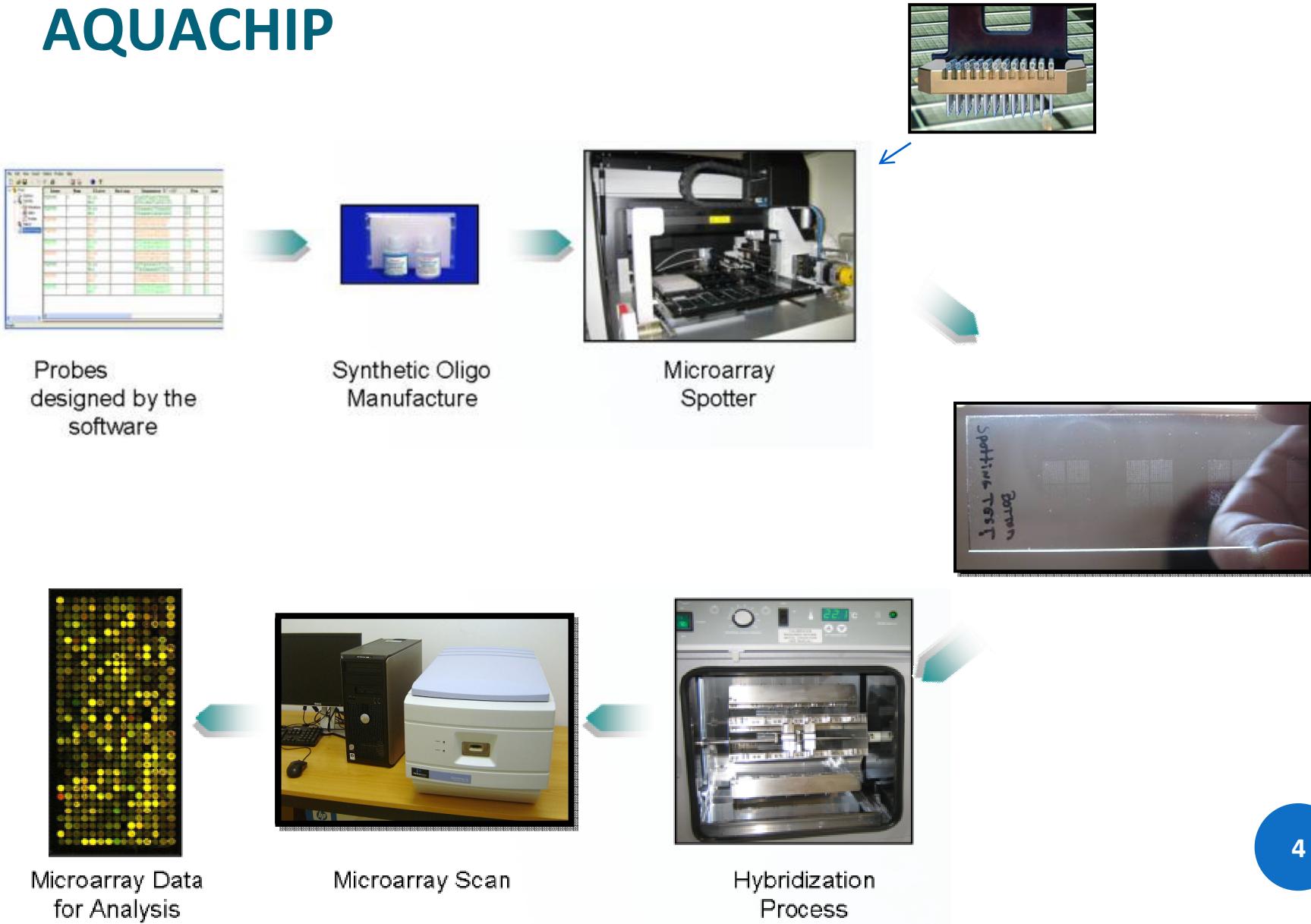
# General concepts



## Chip de DNA



# AQUACHIP

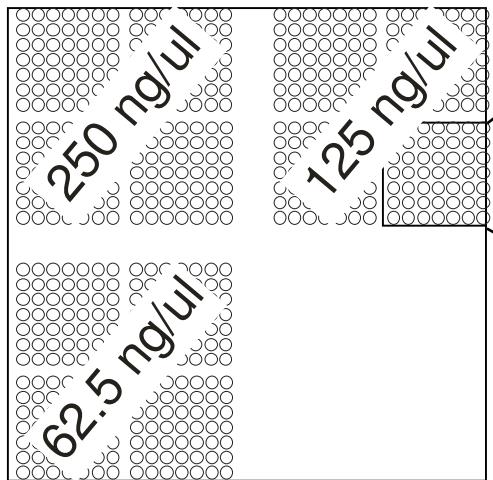


# AQUACHIP

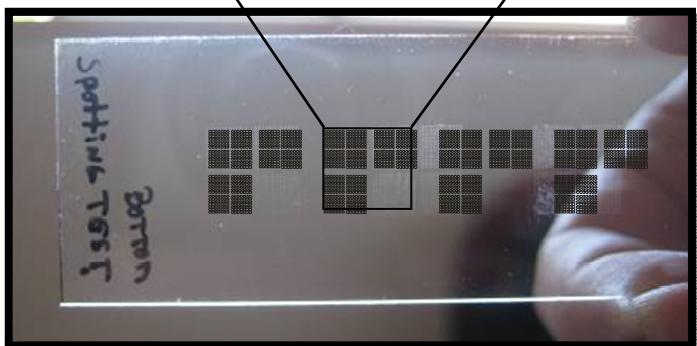
Probe	Microorganism	Sequence	bp
Microorganisms	1	16S rRNA	147
	2	23S rRNA	140
	3	23S rRNA	161
	4	putative toxin of gyrase inhibiting toxin-antitoxin system (locus_tag "ECP_0049")	162
	5	<i>YeeR</i>	186
	6	<i>spoVD</i>	184
	7	<i>uidA</i>	139
	8	<i>lacZ</i>	187
	9	GAP (glyceraldehyde-3-phosphate dehydrogenase, type I)	128
	10	<i>dotA</i>	128
	11	putative_enterotoxin (ECs3855)	152
	12	<i>pebA</i>	107
	13	<i>gyrB</i>	134
	15	<i>ipaH_1</i>	111
	16	<i>hha</i>	164
	17	<i>aroG</i>	110
	18	<i>oprL</i>	141
Control	C1	Negative control	148
	C2	Positive control	148

# AQUACHIP

Random distribution  
3 different probe concentration  
Each probe spotted 8 times at each concentration



Blank	Blank	7	18	17	20	C1
4	2	5	1	C2	10	12
Blank	13	13	6	1	9	2
6	15	9	19	18	C2	11
17	11	12	19	15	8	3
3	Blank	16	7	10	Blank	Blank
20	8	4	5	16	Blank	C1



#### Mandatory species/groups

- 1 – Total coliforms
- 2 – *Enterococcus* spp.
- 3 – *Enterococcus* spp.
- 4 – *E. coli*
- 5 – *E. coli*
- 6 – *C. perfringens*
- 7 – *E. coli*
- 8 – *E. coli*

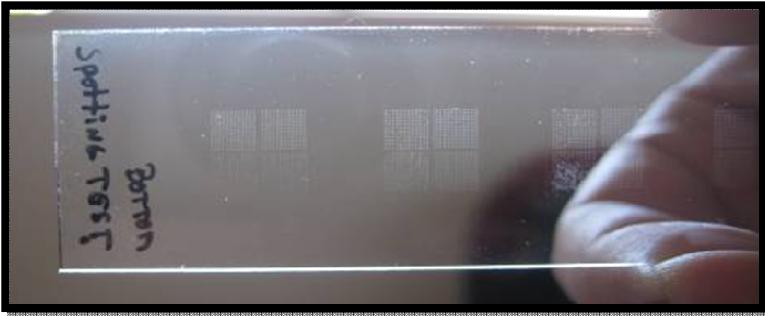
#### Non-mandatory species/groups

- 9 – *S. aureus*
- 10 – *L. pneumophila*
- 11 – *E. coli* O157
- 12 – *Campylobacter* spp.
- 13 – *Campylobacter* spp.
- 15 – *Shigella* spp.
- 16 – Faecal coliforms
- 17 – *Salmonella* spp.
- 18 – *P. aeruginosa*

#### Controls

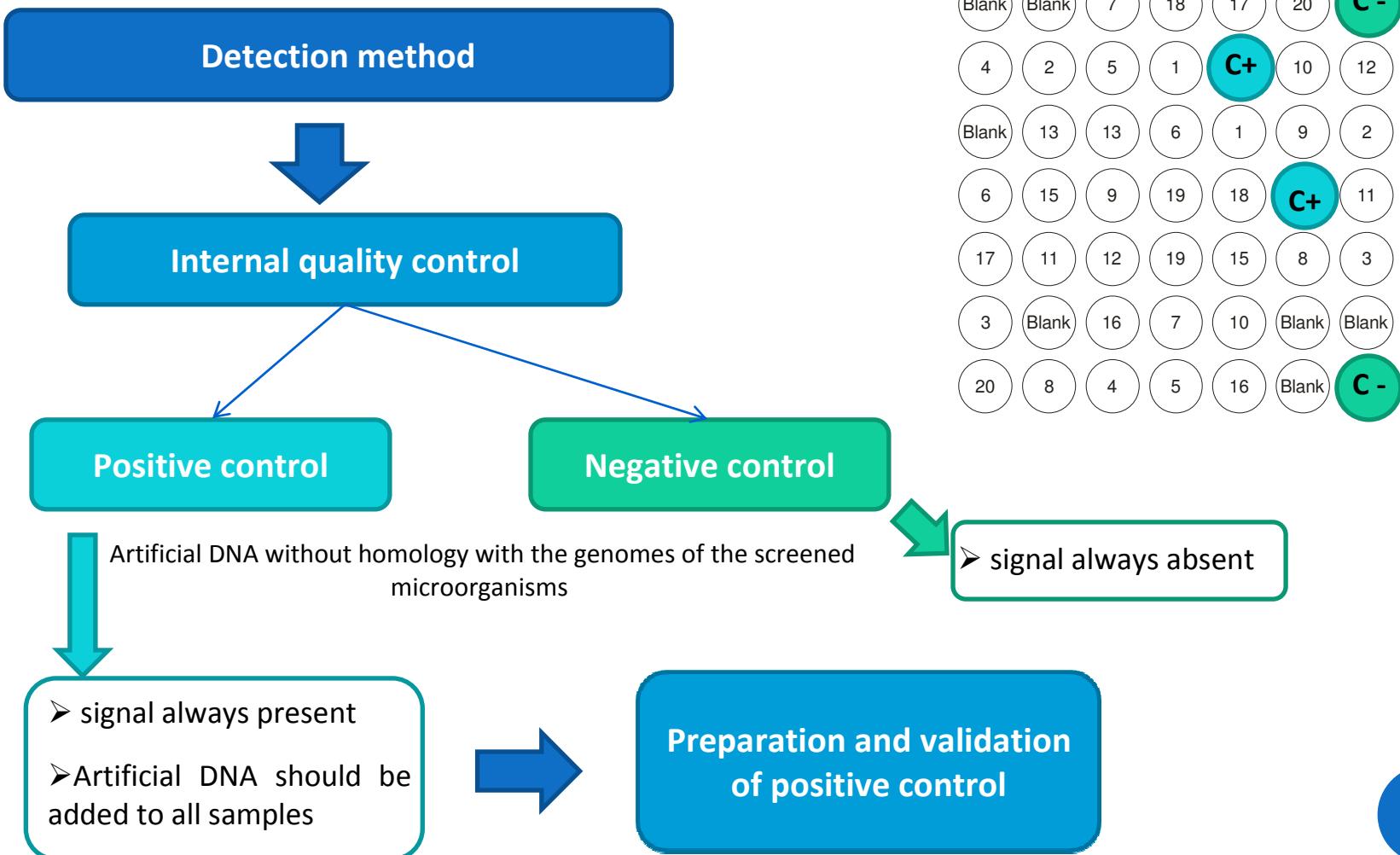
- C1 – Negative control
- C2 – Positive control

## Probe and chip fabrication

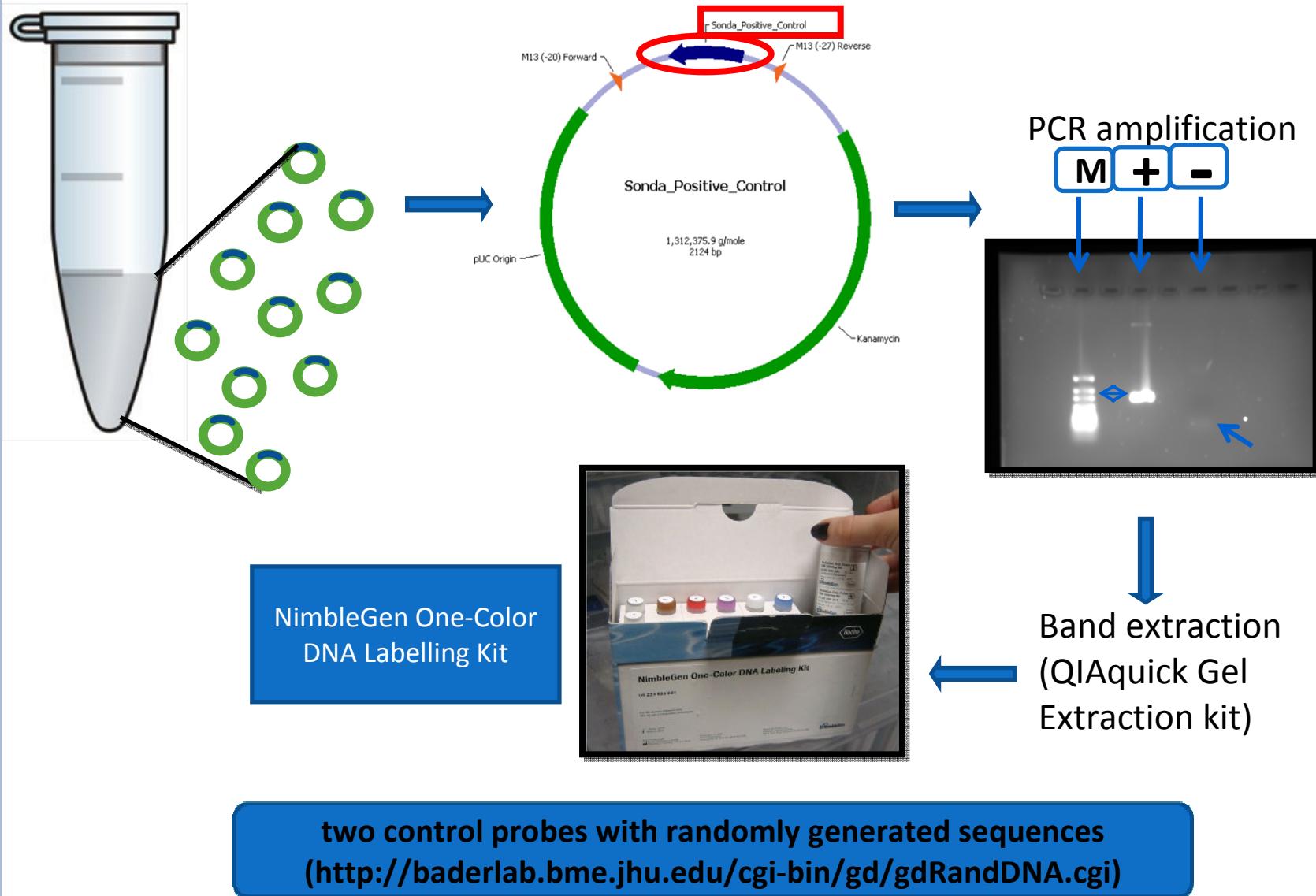


- Single-strand probe synthesis using 3' modified primers with a C6 spacer, pretreatment of Codelink® slides (Surmodics, Eden Prairie, MN, USA).
- Probe spotting, covalent binding of probes to slides and slide washing were all done at the external contractor facilities (Biocant, Portugal).

# 1. Positive control validation



# 1. Positive control preparation



# 1. Positive control hybridization

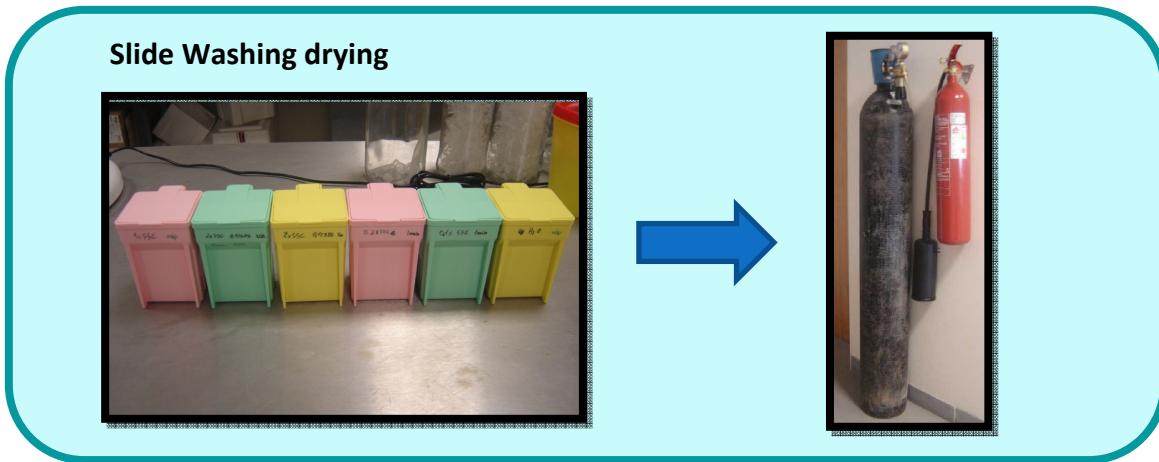
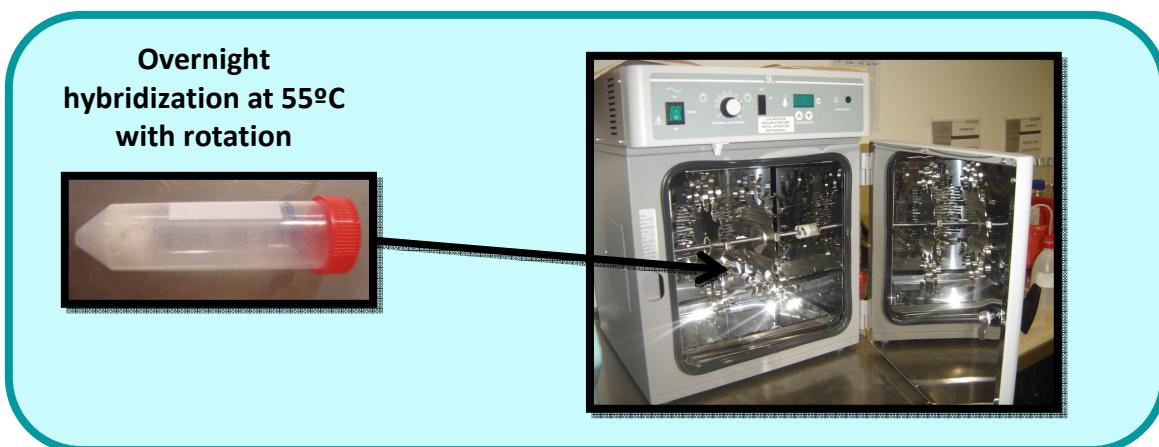
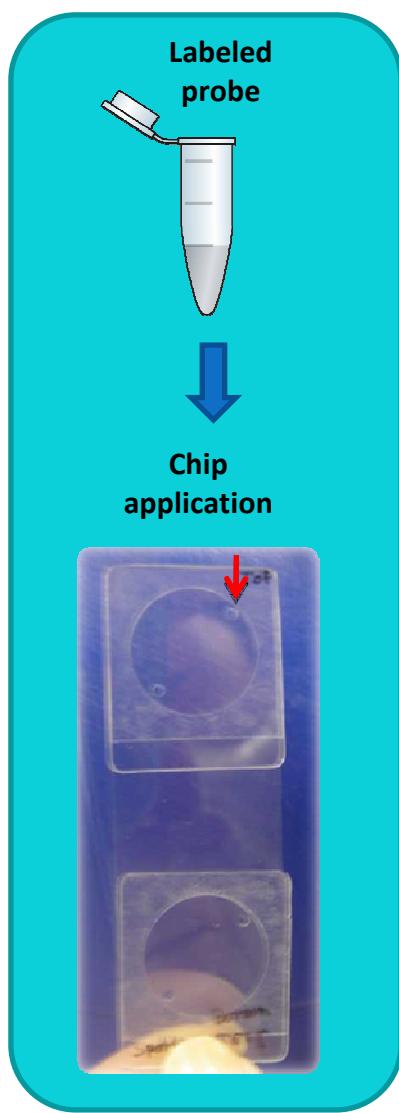
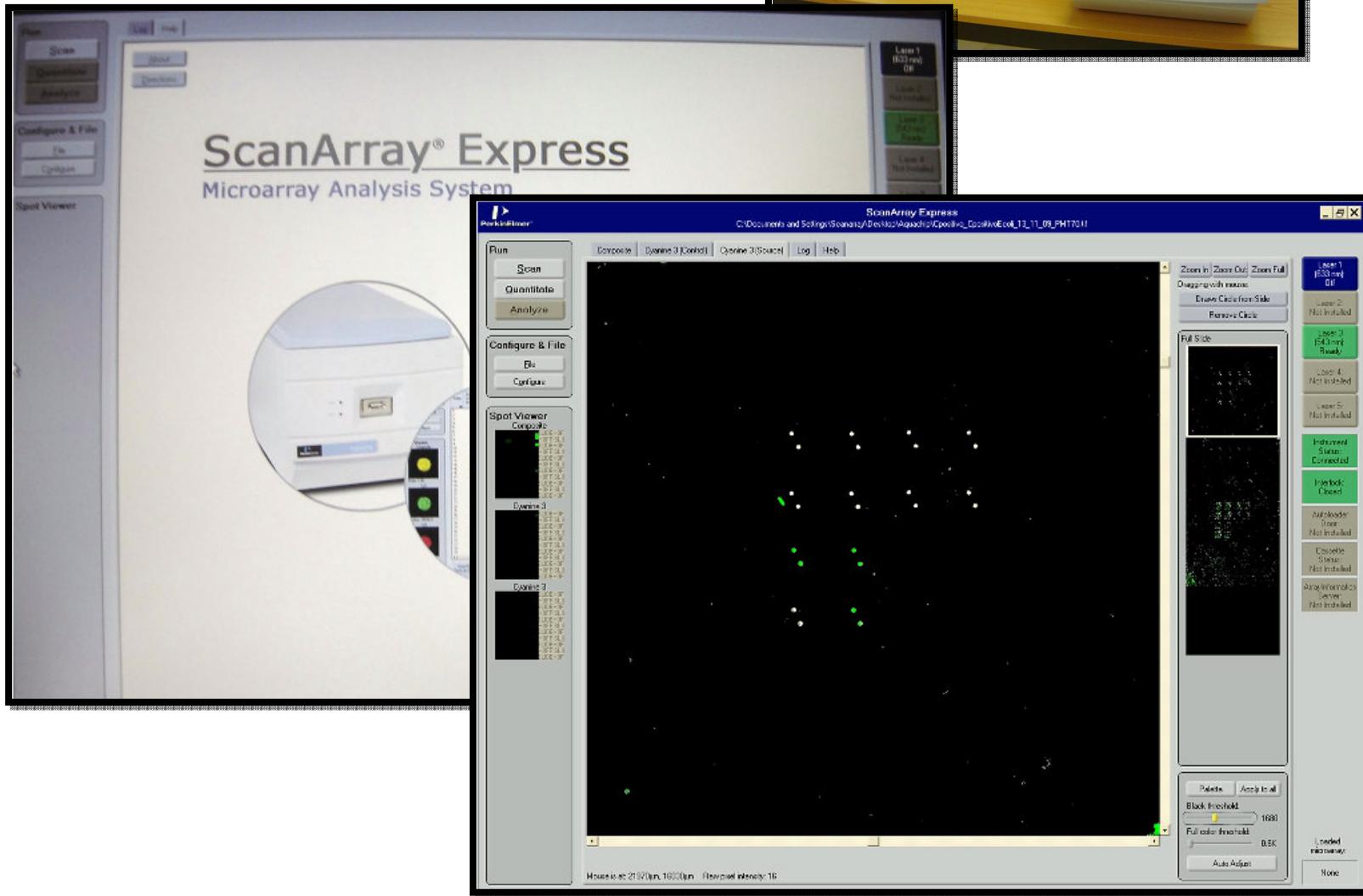


Image acquisition and data analysis

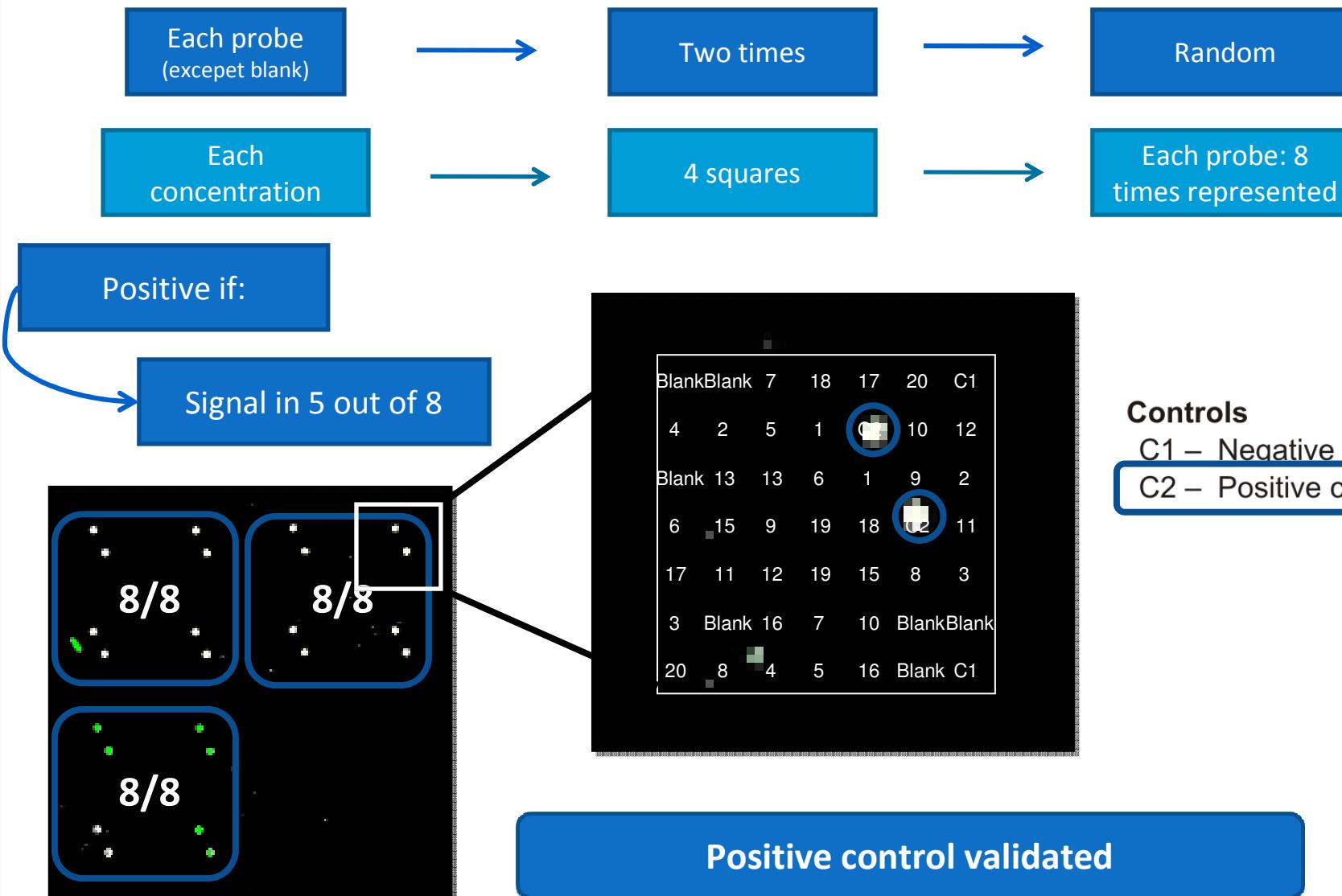
## 1. Positive control validation

### Image acquisition



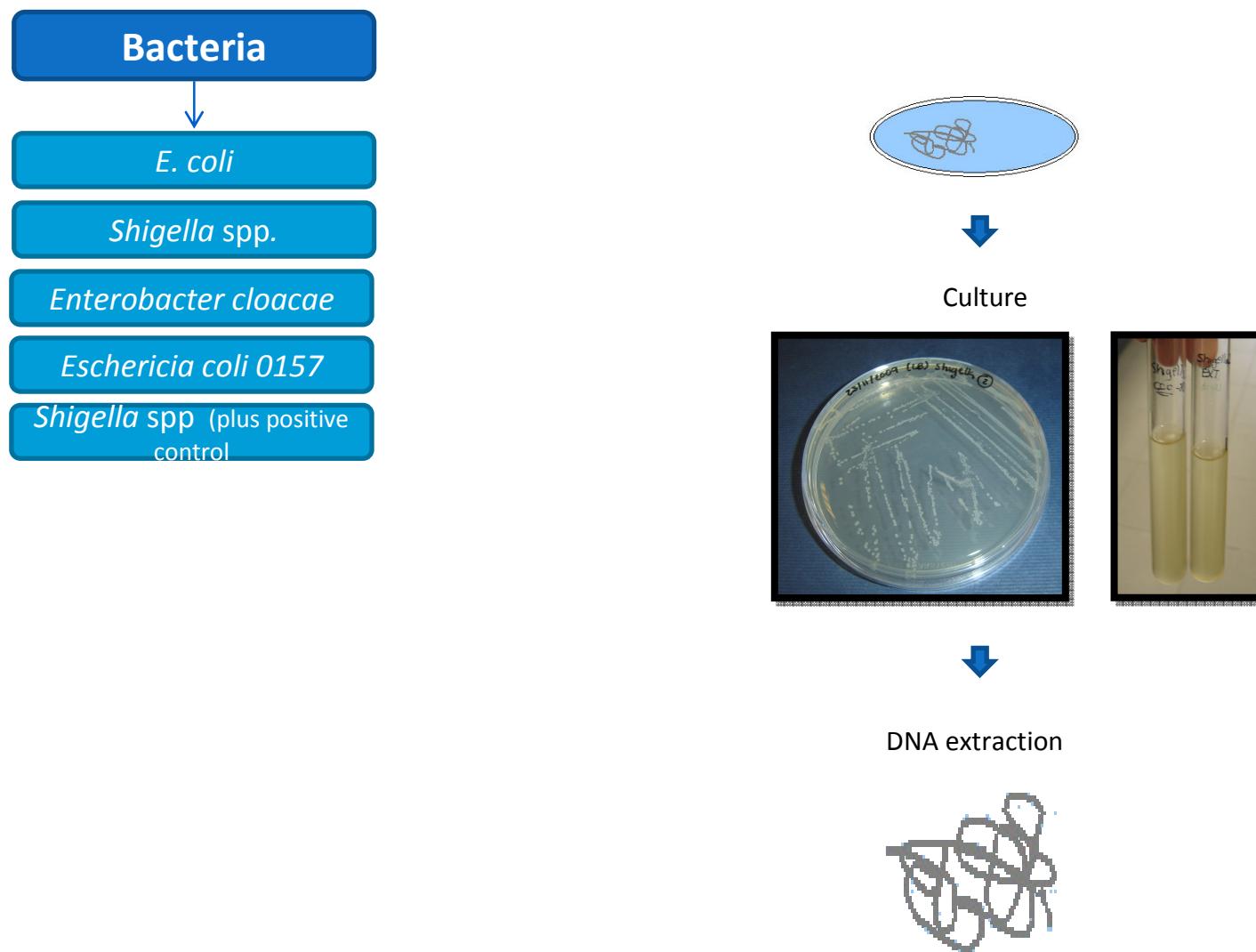
## 1. Positive control validation

### Data analysis



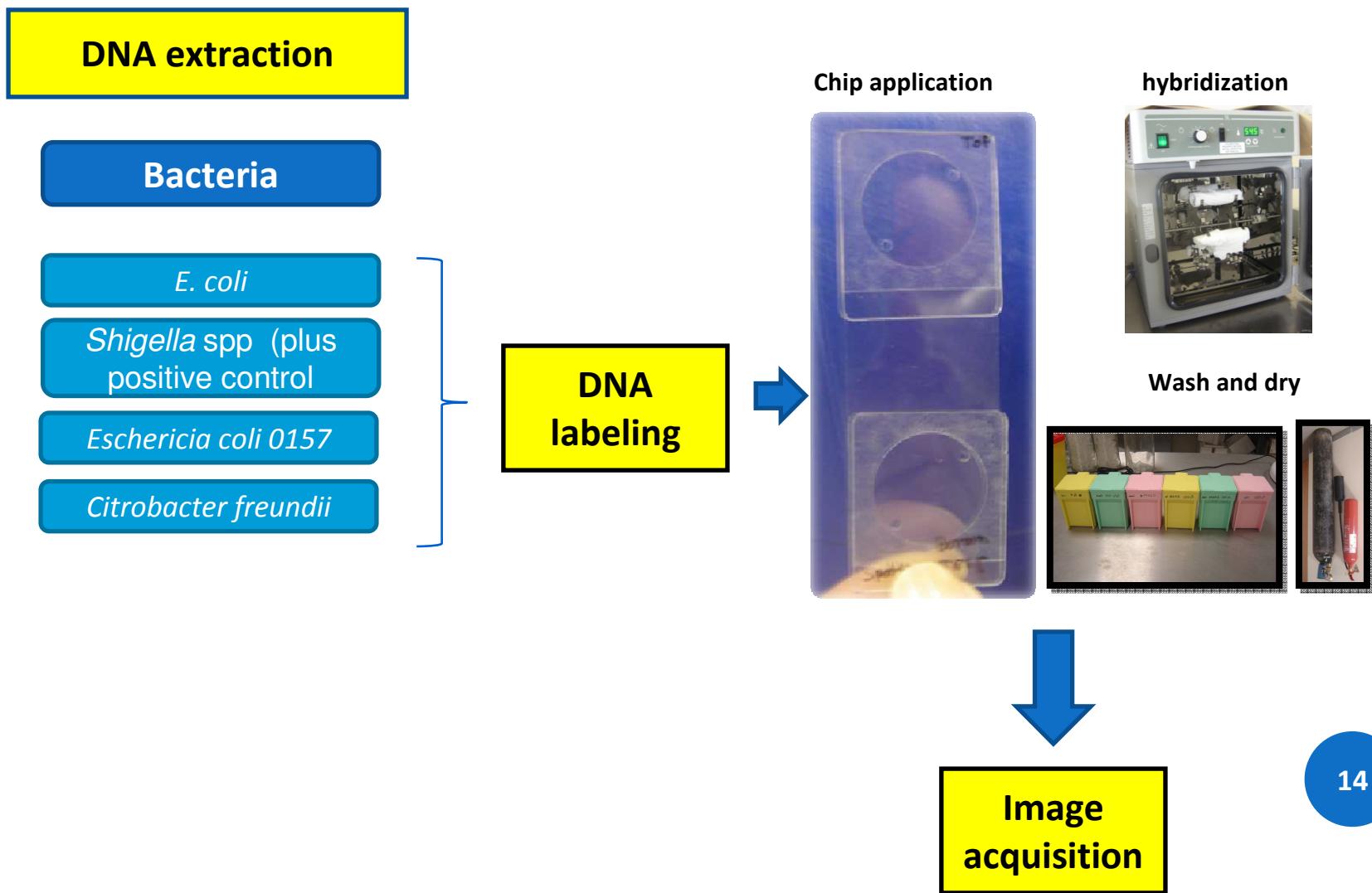
## 2. Validation of other probes

# Sample preparation



## 2. Validation of other probes: some examples

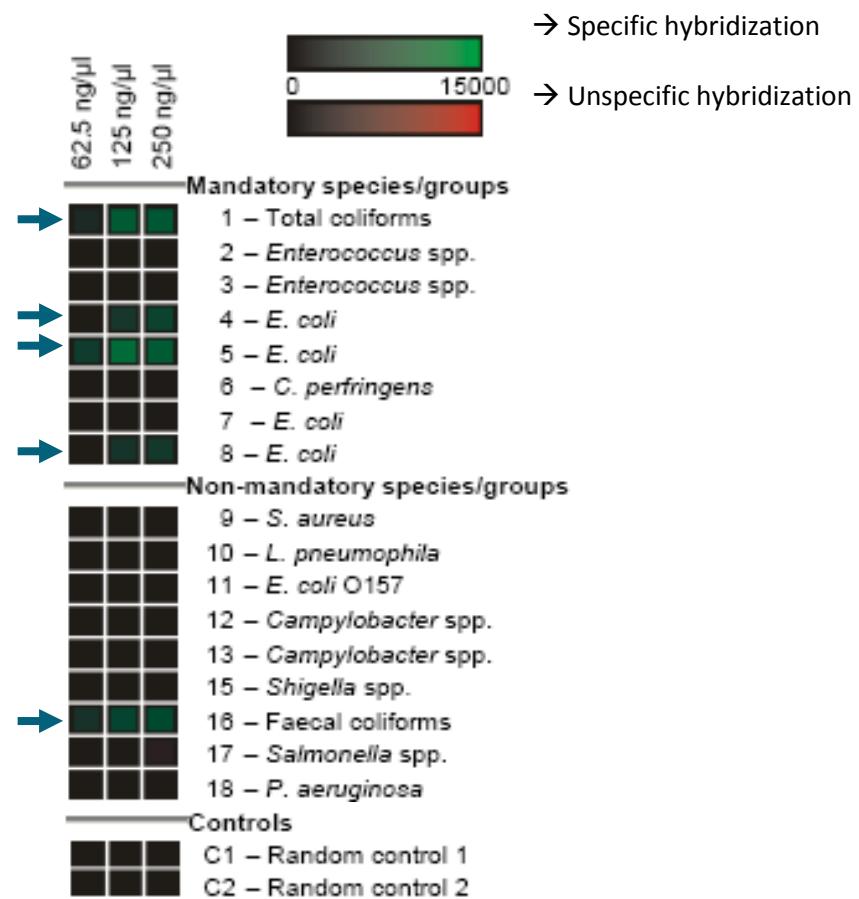
### Labelling and hybridization



## Probes for *E. coli* : validation

➤ Test different probes

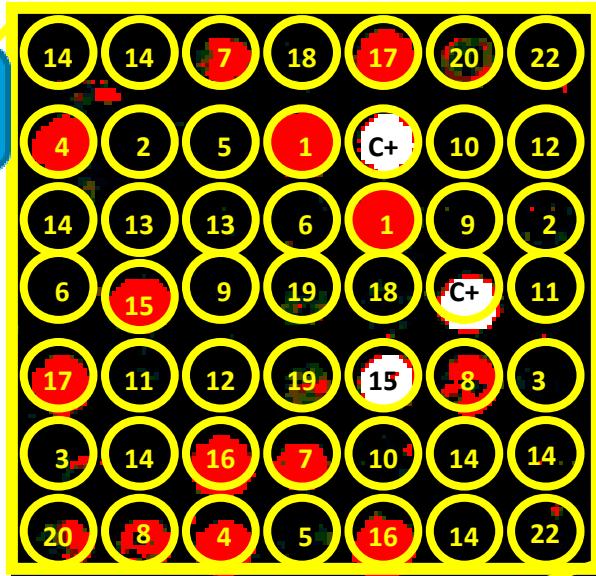
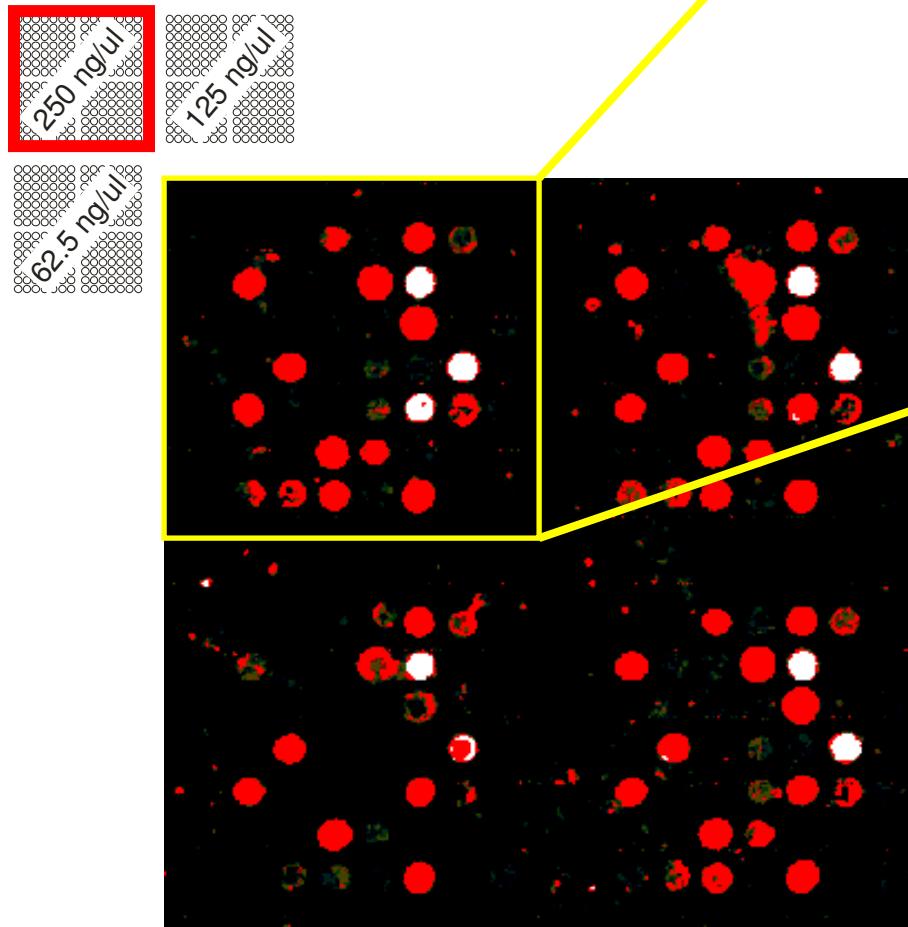
➤ *Escherichia coli*



Probe 4 weak signal

Probe 7 no signal

## *Shigella* spp (plus positive control) : validation

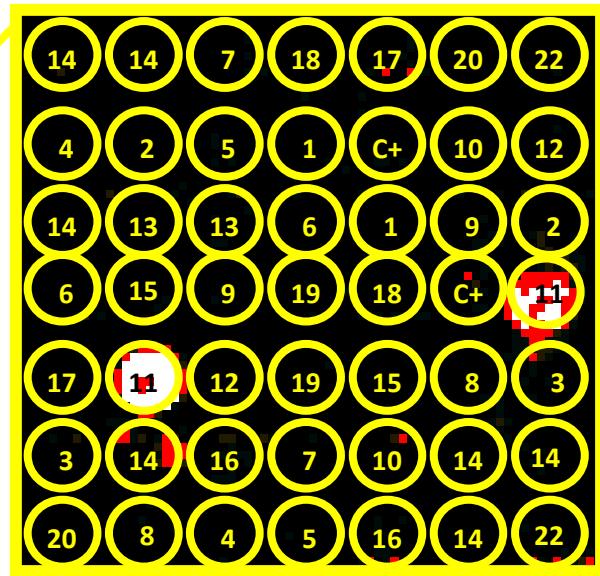
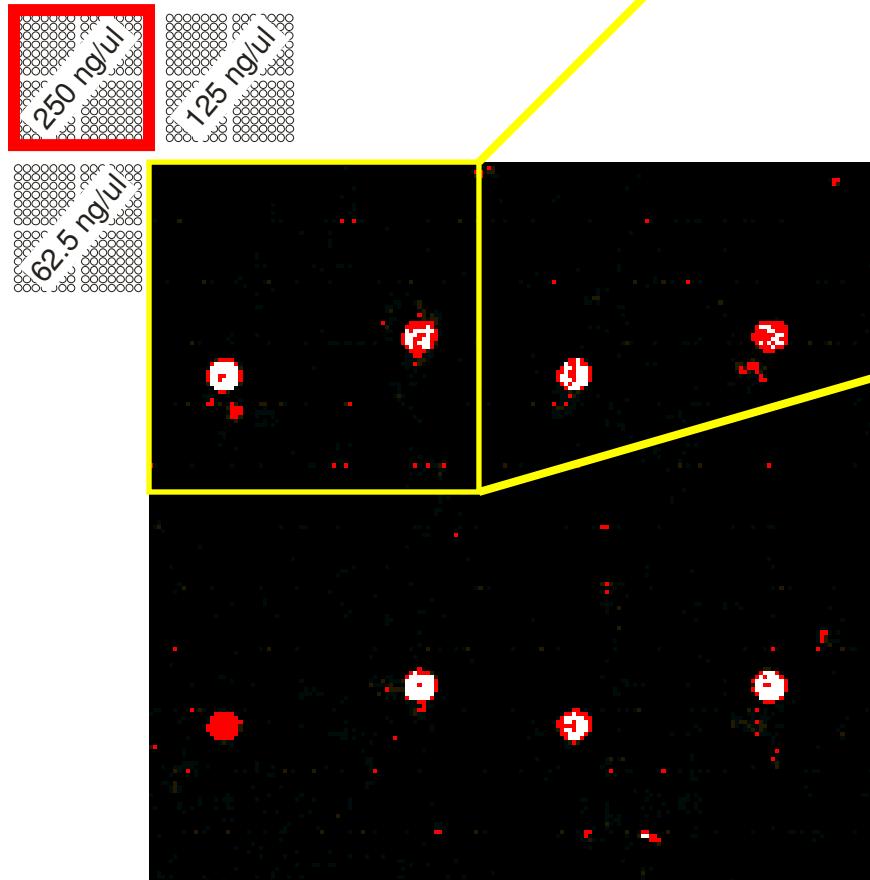


### Results

- 1 – Total coliform
- 16 – Faecal coliform
- 15 - *Shigella*
- C+ - positive control
- 4,7,8 – *E. coli* X
- 17 – *Salmonella* X

16

## *Escherichia coli* O157

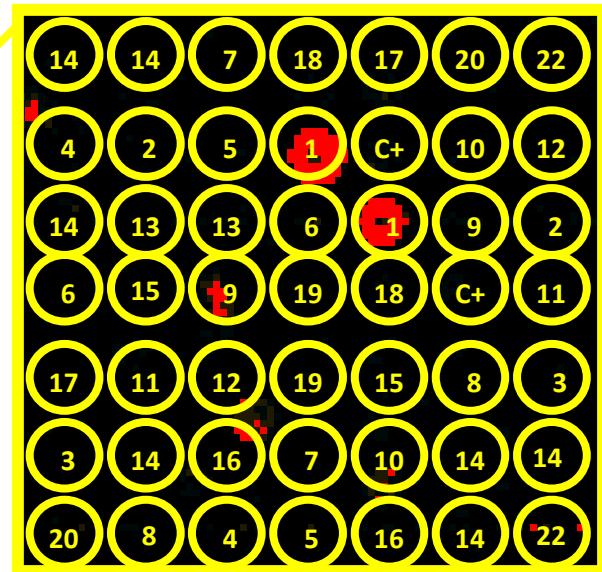
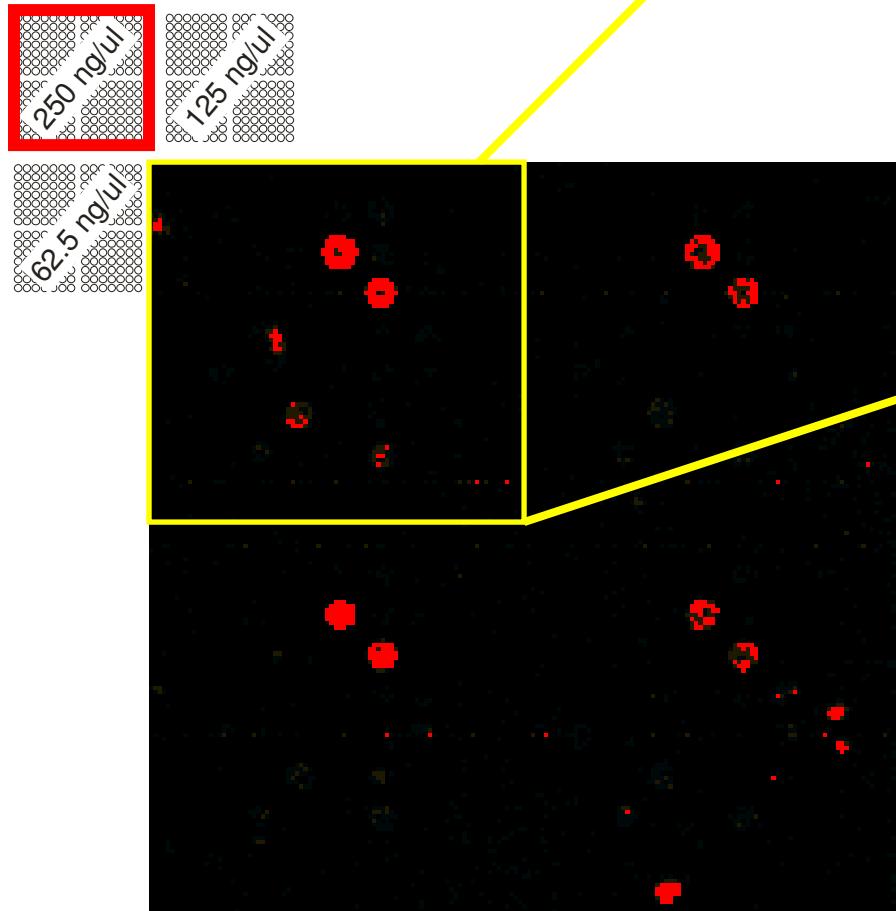


## Results

11 – *E. coli* O157

Without hybridization with  
other *E. coli* probes

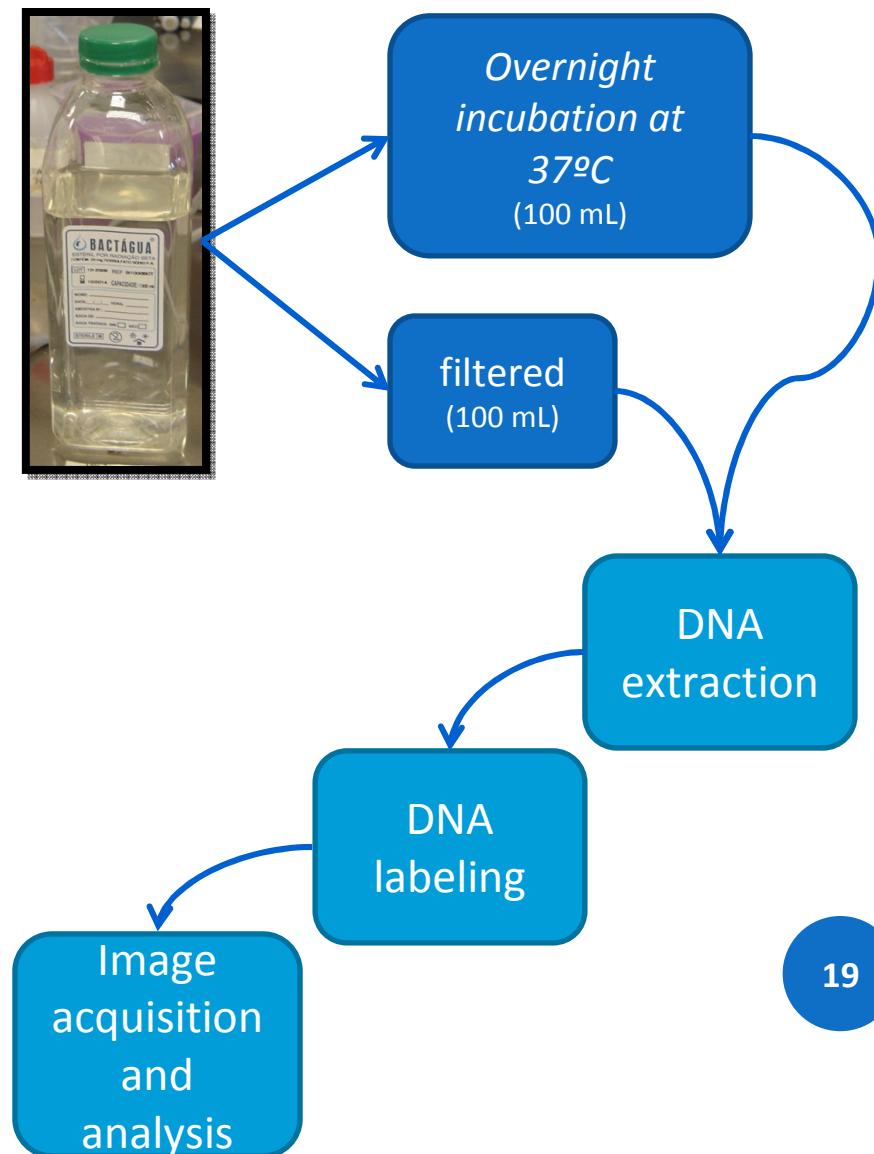
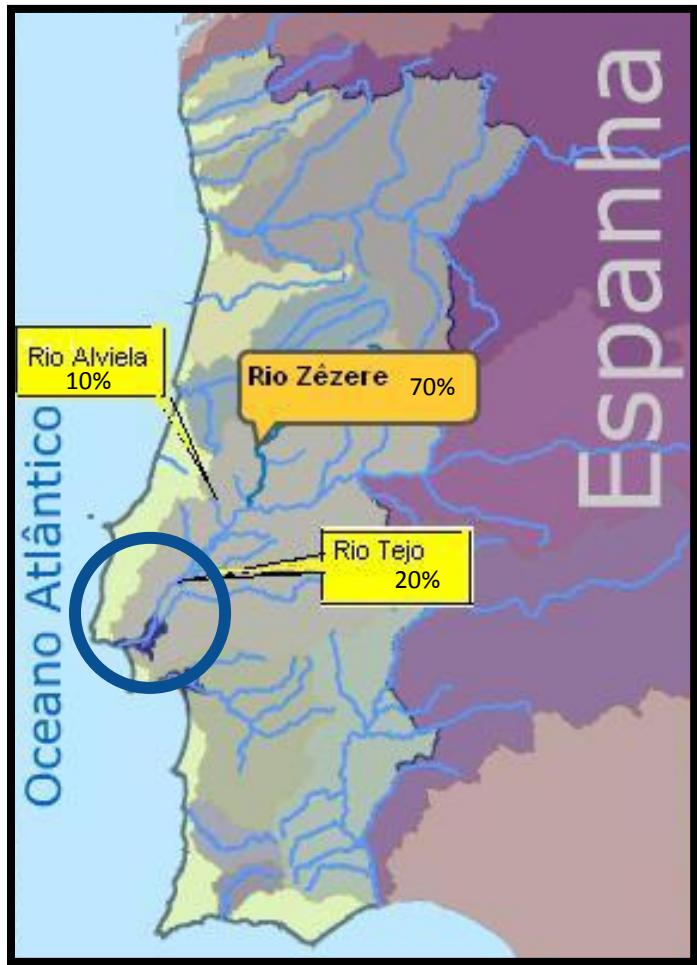
## *Citrobacter freundii*



## Results

1 – Total coliform

### 3. Real water test: Tagus river captation of EPAL (untreated water!)



# Water filtration...



DNA extraction  
from water  
sample

DNA labelling  
and  
hybridization

result

### 3. Real water test: Tagus river captation of EPAL (untreated water!)

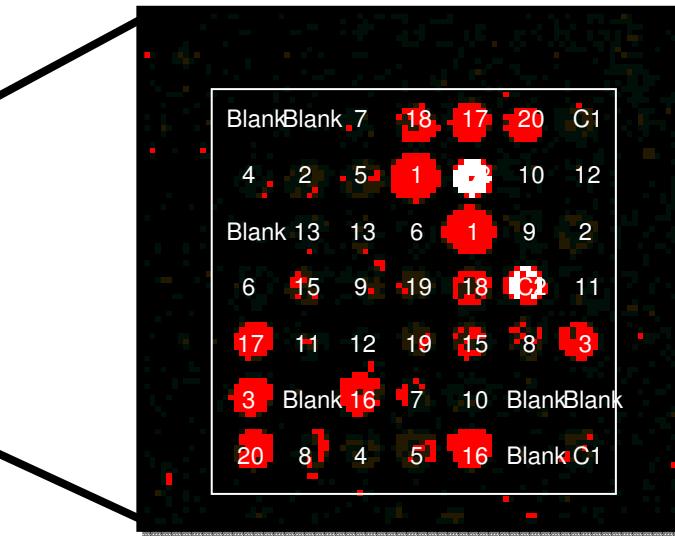
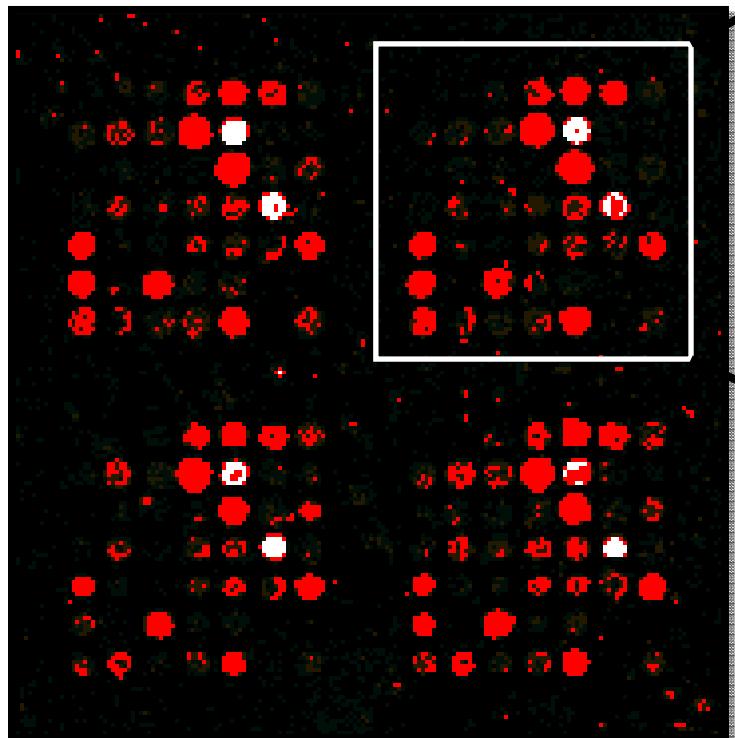
## CHIP RESULTS

Tagus River  
directly filtered

No detection

Culture Tagus River + positive control

250 ng/ $\mu$ L



- C2 – Controlo Positivo      15 – *Shigella* spp.
- 1 – Coliforme Total      16 – Coliforme Fecal
- 2 – *Enterococcus* spp.      17 – *Salmonella* spp.
- 3 - *Enterococcus* spp.      18 – *P.aeruginosa*
- 5 – *E.coli*
- 8 - *E.coli*

## Chip vs. Conventional results

	Probe	Microorganism	Chip positive	ufc/100ml	Conventional method
Microorganisms	1	Total coliforms	✓	2282	Total coliforms
	2	<i>E. faecalis</i>	✓		Enterococcus
	3	<i>E. faecium</i>	✓	68	
	4	<i>E. coli</i>		268	<i>E. coli</i>
	5	<i>E. coli</i>	✓		
	6	<i>C. perfringens</i>			
	7	<i>E. coli</i>			
	8	<i>E. coli</i>	✓		
	9	<i>S. aureus</i>			
	10	<i>L. pneumophila</i>			
	11	<i>E. coli O157</i>			
	12	<i>C. jejuni</i>			
	13	<i>C. coli</i>			
	15	<i>Shigella</i> spp.	✓		
	16	Faecal coliforms	✓	282	Faecal coliforms
	17	<i>Salmonella</i> spp.	✓	Absence 1L	<i>Salmonella</i> spp.
	18	<i>P. aeruginosa</i>	✓	0	<i>P. aeruginosa</i>
Control	C1	Random sequence 1			
	C2	Random sequence 2	✓		

## Chip vs. Conventional results

	Probe	Microorganism	Chip positive	ufc/100ml	Conventional method
Microorganisms	1	Total coliforms	✓	2282	Total coliforms
	2	<i>E. faecalis</i>	✓		Enterococcus
	3	<i>E. faecium</i>	✓	68	
	4	<i>E. coli</i>	✗	268	<i>E. coli</i>
	5	<i>E. coli</i>	✓		
	6	<i>C. perfringens</i>			
	7	<i>E. coli</i>	✗		
	8	<i>E. coli</i>	✓		
	9	<i>S. aureus</i>			
	10	<i>L. pneumophila</i>			
	11	<i>E. coli O157</i>			
	12	<i>C. jejuni</i>			
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	18	<i>P. aeruginosa</i>	✗	0	<i>P. aeruginosa</i>
Control	C1	Random sequence 1			
	C2	Random sequence 2	✓		

# CONCLUSION

- ❖ Positive control validation
  - ❖ Correct hybridization
  - ❖ Helpful for array alignment and identification of positive spots in samples of unknown composition
- ❖ *E. coli* (principal indicator bacteria) with 4 different probes on chip
  - ❖ Two of these probes give expected results (5 and 8) and the other two (4 and 7) should not be considered
- ❖ *E. coli* O157; *Citrobacter freundii*
  - ❖ Correct hybridization
- ❖ Shigella probe
  - ❖ DNA from *Shigella* hybridizes with probes for *Shigella* and *E. coli*
  - ❖ DNA from *E. coli* do not hybridize with probes for *Shigella*
  - ❖ Great homology between *Shigella* spp. and *E. coli* genomes

# CONCLUSION

- ❖ Real water: untreated water from Tagus river captation of EPAL
  - ❖ pre culture step
  - ❖ increase the sample volume of filtered water
  - ❖ discordant results for Pseudomonas spp. and Salmonella spp. (chip positive only)
    - ❖ *Salmonella* probe, which has 95% homology to *E. coli* DNA, according to BLAST [4] (E-value 3E-28), may justify a positive signal
    - ❖ Stressed bacteria may be unable to grow in selective mediums
  - ❖ general agreement of results between Aquachip and conventional methods
  - ❖ reinforces the utility and the proof-of-concept of the DNA chip

## ACKNOWLEDGMENTS

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